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#### (54) Title: GROWTH DIFFERENTIATION FACTOR-9

#### (57) Abstract

Growth differentiation factor-9 (GDF-9) is disclosed along with its polynucleotide sequence and amino acid sequence. Also disclosed are diagnostic and therapeutic methods of using the GDF-9 polypeptide and polynucleotide sequences.

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### **GROWTH DIFFERENTIATION FACTOR-9**

This application is a continuation-in-part application of U.S. Serial No. 08/003,303, filed January 12, 1993.

#### BACKGROUND OF THE INVENTION

#### 5 1. Field of the Invention

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The invention relates generally to growth factors and specifically to a new member of the transforming growth factor beta (TGF- $\beta$ ) superfamily, which is denoted, growth differentiation factor-9 (GDF-9).

#### 2. Description of Related Art

The transforming growth factor *β* (TGF-*β*) superfamily encompasses a group of structurally-related proteins which affect a wide range of differentiation processes during embryonic development. The family includes, Mullerian inhibiting substance (MIS), which is required for normal male sex development (Behringer, *et al.*, *Nature*, 345:167, 1990), *Drosophila* decapentaplegic (DPP) gene product, which is required for dorsal-ventral axis formation and morphogenesis of the imaginal disks (Padgett, *et al.*, *Nature*, 325:81-84, 1987), the *Xenopus* Vg-1 gene product, which localizes to the vegetal pole of eggs ((Weeks, *et al.*, *Cell*, 51:861-867, 1987), the activins (Mason, *et al.*, *Biochem, Biophys. Res. Commun.*, 135:957-964, 1986), which can induce the formation of mesoderm and anterior structures in *Xenopus* embryos (Thomsen, *et al.*, *Cell*, 63:485, 1990), and the bone morphogenetic proteins (BMPs, osteogenin, OP-1) which can induce *de novo* cartilage and bone formation (Sampath, *et al.*, *J. Biol. Chem.*, 265:13198, 1990). The TGF-*β*s can influence a variety of differentiation processes, including adipogenesis, myogenesis, chondrogenesis,

hematopoiesis, and epithelial cell differentiation (for review, see Massague, *Cell* 49:437, 1987).

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The proteins of the TGF- $\beta$  family are initially synthesized as a large precursor protein which subsequently undergoes proteolytic cleavage at a cluster of basic residues approximately 110-140 amino acids from the C-terminus. The C-terminal regions of the proteins are all structurally related and the different family members can be classified into distinct subgroups based on the extent of their homology. Although the homologies within particular subgroups range from 70% to 90% amino acid sequence identity, the homologies between subgroups are significantly lower, generally ranging from only 20% to 50%. In each case, the active species appears to be a disulfide-linked dimer of C-terminal fragments. For most of the family members that have been studied, the homodimeric species has been found to be biologically active, but for other family members, like the inhibins (Ling, et al., Nature, 321:779, 1986) and the TGF- $\beta$ s (Cheifetz, et al., Cell, 48:409, 1987), heterodimers have also been detected, and these appear to have different biological properties than the respective homodimers.

The inhibins and activins were originally purified from follicular fluid and shown to have counteracting effects on the release of follicle-stimulating hormone by the pituitary gland. Although the mRNAs for all three inhibin/activin subunits ( $\alpha a$ ,  $\beta A$  and  $\beta B$ ) have been detected in the ovary, none of these appear to be ovary-specific (Meunier, et al., Proc.Natl.Acad.Sci. USA, 85:247, 1988). MIS has also been shown to be expressed by granulosa cells and the effects of MIS on ovarian development have been documented both *in vivo* in transgenic mice expressing MIS ectopically (Behringer, *supra*) and *in vitro* in organ culture (Vigier, et al., Development, 100:43, 1987).

Identification of new factors that are tissue-specific in their expression pattern will provide a greater understanding of that tissue's development and function.

## SUMMARY OF THE INVENTION

The present invention provides a cell growth and differentiation factor, GDF-9, a polynucleotide sequence which encodes the factor and antibodies which are immunoreactive with the factor. This factor appears to relate to various cell proliferative disorders, especially those involving ovarian tumors, such as granulosa cell tumors.

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Thus, in one embodiment, the invention provides a method for detecting a cell proliferative disorder of ovarian origin and which is associated with GDF-9. In another embodiment, the invention provides a method of treating a cell proliferative disorder associated with abnormal levels of expression of GDF-9, by suppressing or enhancing GDF-9 activity.

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIGURE 1 shows expression of GDF-9 mRNA in adult tissues.

FIGURE 2 shows nucleotide and predicted amino acid sequence of murine GDF-9. Consensus N-glycosylation signals are denoted by plain boxes. The putative tetrabasic processing sites are denoted by stippled boxes. The inframe termination codons upstream of the putative initiating ATG and the consensus polyadenylation signals are underlined. The poly A tails are not shown. Numbers indicate nucleotide position relative to the 5' end.

FIGURE 3 shows the alignment of the C-terminal sequences of GDF-9 with other members of the TGF- $\beta$  family. The conserved cysteine residues are shaded. Dashes denote gaps introduced in order to maximize alignment.

FIGURE 4 shows amino acid homologies among the different members of the TGF- $\beta$  superfamily. Numbers represent percent amino acid identities between each pair calculated from the first conserved cysteine to the C-terminus. Boxes represent homologies among highly-related members within particular subgroups.

FIGURE 5 shows the immunohistochemical localization of GDF-9 protein. Adjacent sections of an adult ovary were either stained with hematoxylin and eosin (FIGURE 5a) or incubated with immune (FIGURE 5b) or pre-immune (FIGURE 5c) serum at a dilution of 1:500. Anti-GDF-9 antiserum was prepared by expressing the C-terminal portion of murine GDF-9 (residues 308-441) in bacteria, excising GDF-9 protein from preparative SDS gels, and immunizing rabbits. Sites of antibody binding were visualized using the Vectastain ABC kit (Vector Labs).

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FIGURE 6 shows a comparison of the predicted amino acid sequences of murine (top lines) and human (bottom lines) GDF-9. Numbers represent amino acid positions relative to the N-termini. Vertical lines represent sequence identities. Dots represent gaps introduced in order to maximize the alignment. The clear box shows the predicted proteolytic processing sites. The shaded boxes show the cysteine residues in the mature region of the proteins. The bars at the bottom show a schematic of the pre-(clear) and mature (shaded) regions of GDF-9 with the percent sequence identities between the murine and human sequences shown below.

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FIGURE 7 shows *in situ* hybridization to adult ovary sections using a GDF-9 RNA probe. [<sup>35</sup>S]-labeled anti-sense (FIGURE 7a and 7c) or sense (FIGURE 7 b and 7d) GDF-9 RNA probes were hybridized to adjacent paraffinembedded sections of ovaries fixed in 4% paraformaldehyde. Sections were dipped in photographic emulsion, exposed, developed, and then stained with hematoxylin and eosin. Two representative fields are shown.

FIGURE 8 shows *in situ* hybridization to a postnatal day 4 ovary section using an antisense GDF-9 RNA probe. Sections were prepared as described for FIGURE 7. Following autoradiography and staining, the section was photographed under bright-field (FIGURE 8a) or dark-field (FIGURE 8b) illumination.

FIGURE 9 shows *in situ* hybridization to postnatal day 8 ovary sections using an antisense (FIGURE 9a) or sense (FIGURE 9b) GDF-9 RNA probe. Sections were prepared as described for FIGURE 7.

FIGURE 10 shows *in situ* hybridization to adult oviduct sections using an antisense (FIGURE 10a) or sense (FIGURE 10b) GDF-9 RNA probe. Sections were prepared as described for FIGURE 7.

FIGURE 11 shows *in situ* hybridization to an adult oviduct (0.5 days following fertilization) section using an antisense GDF-9 RNA probe. Sections were prepared as described for FIGURE 7. Following autoradiography and staining, the section was photographed under bright-field (FIGURE 11a) or dark-field (FIGURE 11b) illumination.

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#### DETAILED DESCRIPTION OF THE INVENTION

The present invention provides a growth and differentiation factor, GDF-9 and a polynucleotide sequence encoding GDF-9. Unlike other members of the TGF- $\beta$  superfamily, GDF-9 expression is highly tissue specific, being expressed in cells primarily in ovarian tissue. In one embodiment, the invention provides a method for detection of a cell proliferative disorder of the ovary, which is associated with GDF-9 expression. In another embodiment, the invention provides a method for treating a cell proliferative disorder associated with abnormal expression of GDF-9 by using an agent which suppresses or enhances GDF-9 activity.

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The TGF- $\beta$  superfamily consists of multifunctionaly polypeptides that control proliferation, differentiation, and other functions in many cell types. Many of the peptides have regulatory, both positive and negative, effects on other peptide growth factors. The structural homology between the GDF-9 protein of this invention and the members of the TGF- $\beta$  family, indicates that GDF-9 is a new member of the family of growth and differentiation factors. Based on the known activities of many of the other members, it can be expected that GDF-9 will also possess biological activities that will make it useful as a diagnostic and therapeutic reagent.

For example, another regulatory protein that has been found to have structural homology with TGF- $\beta$  is inhibin, a specific and potent polypeptide inhibitor of the pituitary secretion of FSH. Inhibin has been isolated from ovarian follicular fluid. Because of its suppression of FSH, inhibin has potential to be used as a contraceptive in both males and females. GDF-9 may possess similar biological activity since it is also an ovarian specific peptide.Inhibin has also been shown to be useful as a marker for certain ovarian tumors (Lappohn, et al., N. Engl. J. Med., 321:790, 1989). GDF-9 may also be useful as a marker

for identifying primary and metastatic neoplasms of ovarian origin. Similarly, GDF-9 may be useful as an indicator of developmental anomalies in prenatal screening procedures.

Another peptide of the TGF- $\beta$  family is MIS, produced by the testis and responsible for the regression of the Mullerian ducts in the male embryo. MIS has been show to inhibit the growth of human ovarian cancer in nude mice (Donahoe, et al., Ann. Surg., 194:472, 1981). GDF-9 may function similarly and may, therefore, be useful as an anti-cancer agent, such as for the treatment of ovarian cancer.

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GDF-9 may also function as a growth stimulatory factor and, therefore, be useful for the survival of various cell populations *in vitro*. In particular, if GDF-9 plays a role in oocyte maturation, it may be useful in *in vitro* fertilization procedures, e.g., in enhancing the success rate. Many of the members of the TGF-β family are also important mediators of tissue repair. TGF-β has been shown to have marked effects on the formation of collagen and causes a striking angiogenic response in the newborn mouse (Roberts, *et al.*, *Proc. Natl. Acad. Sci. USA*, <u>83</u>:4167, 1986). GDF-9 may also have similar activities and may be useful in repair of tissue injury caused by trauma or burns for example.

The term "substantially pure" as used herein refers to GDF-9 which is substantially free of other proteins, lipids, carbohydrates or other materials with which it is naturally associated. One skilled in the art can purify GDF-9 using standard techniques for protein purification. The substantially pure polypeptide will yield a single major band on a non-reducing polyacrylamide gel. The purity of the GDF-9 polypeptide can also be determined by amino-terminal amino acid sequence analysis. GDF-9 polypeptide includes functional fragments of the polypeptide, as long as the activity of GDF-9 remains. Smaller peptides containing the biological activity of GDF-9 are included in the invention.

The invention provides polynucleotides encoding the GDF-9 protein. These polynucleotides include DNA, cDNA and RNA sequences which encode GDF-9. It is understood that all polynucleotides encoding all or a portion of GDF-9 are also included herein, as long as they encode a polypeptide with GDF-9 activity. Such polynucleotides include naturally occurring, synthetic, and intentionally manipulated polynucleotides. For example, GDF-9 polynucleotide may be subjected to site-directed mutagenesis. The polynucleotide sequence for GDF-9 also includes antisense sequences. The polynucleotides of the invention include sequences that are degenerate as a result of the genetic code. There are 20 natural amino acids, most of which are specified by more than one codon. Therefore, all degenerate nucleotide sequences are included in the invention as long as the amino acid sequence of GDF-9 polypeptide encoded by the nucleotide sequence is functionally unchanged.

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Specifically disclosed herein is a cDNA sequence for GDF-9 which is 1712 base pairs in length and contains an open reading frame beginning with a methionine codon at nucleotide 29. The encoded polypeptide is 441 amino acids in length with a molecular weight of about 49.6 kD, as determined by The GDF-9 sequence contains a core of nucleotide sequence analysis. hydrophobic amino acids near the N-terminus, suggestive of a signal sequence for secretion. GDF-9 contains four potential N-glycosylation sites at asparagine residues 163, 229, 258, and 325 and a putative tetrabasic proteolytic processing site (RRRR) at amino acids 303-306. The mature C-terminal fragment of GDF-9 is predicted to be 135 amino acids in length and have an unglycosylated molecular weight of about 15.6 kD, as determined by nucleotide sequence analysis. One skilled in the art can modify, or partially or completely remove the glycosyl groups from the GDF-9 protein using standard techniques. Therefore, the functional protein or fragments thereof of the invention includes glycosylated, partially glycosylated and unglycosylated species of GDF-9.

The degree of sequence identity of GDF-9 with known TGF- $\beta$  family members ranges from a minimum of 21% with Mullerian inhibiting substance (MIS) to a maximum of 34% with bone morphogenetic protein-4 (BMP-4). GDF-9 specifically disclosed herein differs from the known family members in its pattern of cysteine residues in the C-terminal region. GDF-9 lacks the fourth cysteine of the seven cysteines present in other family members; in place of cysteine at this position, the GDF-9 sequence contains a serine residue. This GDF-9 does not contain a seventh cysteine residue elsewhere in the C-terminal region.

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Minor modifications of the recombinant GDF-9 primary amino acid sequence may result in proteins which have substantially equivalent activity as compared to the GDF-9 polypeptide described herein. Such modifications may be deliberate, as by site-directed mutagenesis, or may be spontaneous. All of the polypeptides produced by these modifications are included herein as long as the biological activity of GDF-9 still exists. Further, deletion of one or more amino acids can also result in a modification of the structure of the resultant molecule without significantly altering its biological activity. This can lead to the development of a smaller active molecule which would have broader utility. For example, one can remove amino or carboxy terminal amino acids which are not required for GDF-9 biological activity.

The nucleotide sequence encoding the GDF-9 polypeptide of the invention includes the disclosed sequence and conservative variations thereof. The term "conservative variation" as used herein denotes the replacement of an amino acid residue by another, biologically similar residue. Examples of conservative variations include the substitution of one hydrophobic residue such as isoleucine, valine, leucine or methionine for another, or the substitution of one polar residue for another, such as the substitution of arginine for lysine, glutamic for aspartic acids, or glutamine for asparagine, and the like. The term

"conservative variation" also includes the use of a substituted amino acid in place of an unsubstituted parent amino acid provided that antibodies raised to the substituted polypeptide also immunoreact with the unsubstituted polypeptide.

DNA sequences of the invention can be obtained by several methods. For example, the DNA can be isolated using hybridization techniques which are well known in the art. These include, but are not limited to: 1) hybridization of genomic or cDNA libraries with probes to detect homologous nucleotide sequences and 2) antibody screening of expression libraries to detect cloned DNA fragments with shared structural features.

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Preferably the GDF-9 polynucleotide of the invention is derived from a mammalian organism, and most preferably from a mouse, rat, or human. Screening procedures which rely on nucleic acid hybridization make it possible to isolate any gene sequence from any organism, provided the appropriate probe is available. Oligonucleotide probes, which correspond to a part of the sequence encoding the protein in question, can be synthesized chemically. This requires that short, oligopeptide stretches of amino acid sequence must be known. The DNA sequence encoding the protein can be deduced from the genetic code, however, the degeneracy of the code must be taken into It is possible to perform a mixed addition reaction when the sequence is degenerate. This includes a heterogeneous mixture of denatured double-stranded DNA. For such screening, hybridization is preferably performed on either single-stranded DNA or denatured double-stranded DNA. Hybridization is particularly useful in the detection of cDNA clones derived from sources where an extremely low amount of mRNA sequences relating to the polypeptide of interest are present. In other words, by using stringent hybridization conditions directed to avoid non-specific binding, it is possible, for example, to allow the autoradiographic visualization of a specific cDNA clone by the hybridization of the target DNA to that single probe in the mixture which is its complete complement (Wallace, et al., Nucl. Acid Res., 9:879, 1981).

The development of specific DNA sequences encoding GDF-9 can also be obtained by: 1) isolation of double-stranded DNA sequences from the genomic DNA; 2) chemical manufacture of a DNA sequence to provide the necessary codons for the polypeptide of interest; and 3) *in vitro* synthesis of a double-stranded DNA sequence by reverse transcription of mRNA isolated from a eukaryotic donor cell. In the latter case, a double-stranded DNA complement of mRNA is eventually formed which is generally referred to as cDNA.

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Of the three above-noted methods for developing specific DNA sequences for use in recombinant procedures, the isolation of genomic DNA isolates is the least common. This is especially true when it is desirable to obtain the microbial expression of mammalian polypeptides due to the presence of introns.

The synthesis of DNA sequences is frequently the method of choice when the entire sequence of amino acid residues of the desired polypeptide product is known. When the entire sequence of amino acid residues of the desired polypeptide is not known, the direct synthesis of DNA sequences is not possible and the method of choice is the synthesis of cDNA sequences. Among the standard procedures for isolating cDNA sequences of interest is the formation of plasmid- or phage-carrying cDNA libraries which are derived from reverse transcription of mRNA which is abundant in donor cells that have a high level of genetic expression. When used in combination with polymerase chain reaction technology, even rare expression products can be cloned. In those cases where significant portions of the amino acid sequence of the polypeptide are known, the production of labeled single or double-stranded

DNA or RNA probe sequences duplicating a sequence putatively present in the target cDNA may be employed in DNA/DNA hybridization procedures which are carried out on cloned copies of the cDNA which have been denatured into a single-stranded form (Jay, et al., Nucl. Acid Res., 11:2325, 1983).

A cDNA expression library, such as lambda gt11, can be screened indirectly for GDF-9 peptides having at least one epitope, using antibodies specific for GDF-9. Such antibodies can be either polyclonally or monoclonally derived and used to detect expression product indicative of the presence of GDF-9 cDNA.

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DNA sequences encoding GDF-9 can be expressed *in vitro* by DNA transfer into a suitable host cell. "Host cells" are cells in which a vector can be propagated and its DNA expressed. The term also includes any progeny of the subject host cell. It is understood that all progeny may not be identical to the parental cell since there may be mutations that occur during replication. However, such progeny are included when the term "host cell" is used. Methods of stable transfer, meaning that the foreign DNA is continuously maintained in the host, are known in the art.

In the present invention, the GDF-9 polynucleotide sequences may be inserted into a recombinant expression vector. The term "recombinant expression vector" refers to a plasmid, virus or other vehicle known in the art that has been manipulated by insertion or incorporation of the GDF-9 genetic sequences. Such expression vectors contain a promoter sequence which facilitates the efficient transcription of the inserted genetic sequence of the host. The expression vector typically contains an origin of replication, a promoter, as well as specific genes which allow phenotypic selection of the transformed cells. Vectors suitable for use in the present invention include, but are not limited to the T7-based expression vector for expression in bacteria (Rosenberg, et al.,

Gene ,56:125, 1987), the pMSXND expression vector for expression in mammalian cells (Lee and Nathans, *J. Biol. Chem.*, 263:3521, 1988) and baculovirus-derived vectors for expression in insect cells. The DNA segment can be present in the vector operably linked to regulatory elements, for example, a promoter (e.g., T7, metallothionein I, or polyhedrin promoters).

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Polynucleotide sequences encoding GDF-9 can be expressed in either prokaryotes or eukaryotes. Hosts can include microbial, yeast, insect and mammalian organisms. Methods of expressing DNA sequences having eukaryotic or viral sequences in prokaryotes are well known in the art. Biologically functional viral and plasmid DNA vectors capable of expression and replication in a host are known in the art. Such vectors are used to incorporate DNA sequences of the invention.

Transformation of a host cell with recombinant DNA may be carried out by conventional techniques as are well known to those skilled in the art. Where the host is prokaryotic, such as *E. coli*, competent cells which are capable of DNA uptake can be prepared from cells harvested after exponential growth phase and subsequently treated by the CaCl<sub>2</sub> method using procedures well known in the art. Alternatively, MgCl<sub>2</sub> or RbCl can be used. Transformation can also be performed after forming a protoplast of the host cell if desired.

When the host is a eukaryote, such methods of transfection of DNA as calcium phosphate co-precipitates, conventional mechanical procedures such as microinjection, electroporation, insertion of a plasmid encased in liposomes, or virus vectors may be used. Eukaryotic cells can also be cotransformed with DNA sequences encoding the GDF-9 of the invention, and a second foreign DNA molecule encoding a selectable phenotype, such as the herpes simplex thymidine kinase gene. Another method is to use a eukaryotic viral vector, such as simian virus 40 (SV40) or bovine papilloma virus, to transiently infect

or transform eukaryotic cells and express the protein. (see for example, Eukaryotic Viral Vectors, Cold Spring Harbor Laboratory, Gluzman ed., 1982).

Isolation and purification of microbial expressed polypeptide, or fragments thereof, provided by the invention, may be carried out by conventional means including preparative chromatography and immunological separations involving monoclonal or polyclonal antibodies.

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The invention includes antibodies immunoreactive with GDF-9 polypeptide or functional fragments thereof. Antibody which consists essentially of pooled monoclonal antibodies with different epitopic specificities, as well as distinct monoclonal antibody preparations are provided. Monoclonal antibodies are made from antigen containing fragments of the protein by methods well known to those skilled in the art (Kohler, et al., Nature, 256:495, 1975). The term antibody as used in this invention is meant to include intact molecules as well as fragments thereof, such as Fab and F(ab')<sub>2</sub>, which are capable of binding an epitopic determinant on GDF-9.

The term "cell-proliferative disorder" denotes malignant as well as non-malignant cell populations which often appear to differ from the surrounding tissue both morphologically and genotypically. The GDF-9 polynucleotide that is an antisense molecule is useful in treating malignancies of the various organ systems, particularly, for example, the ovaries. Essentially, any disorder which is etiologically linked to altered expression of GDF-9 could be considered susceptible to treatment with a GDF-9 suppressing reagent.

The invention provides a method for detecting a cell proliferative disorder of the ovary which comprises contacting an anti-GDF-9 antibody with a cell suspected of having a GDF-9 associated disorder and detecting binding to the antibody. The antibody reactive with GDF-9 is labeled with a compound which allows

detection of binding to GDF-9. For purposes of the invention, an antibody specific for GDF-9 polypeptide may be used to detect the level of GDF-9 in biological fluids and tissues. Any specimen containing a detectable amount of antigen can be used. A preferred sample of this invention is tissue of ovarian origin, specifically tissue containing granulosa cells or ovarian follicular fluid. The level of GDF-9 in the suspect cell can be compared with the level in a normal cell to determine whether the subject has a GDF-9-associated cell proliferative disorder. Preferably the subject is human.

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The antibodies of the invention can be used in any subject in which it is desirable to administer *in vitro* or *in vivo* immunodiagnosis or immunotherapy. The antibodies of the invention are suited for use, for example, in immunoassays in which they can be utilized in liquid phase or bound to a solid phase carrier. In addition, the antibodies in these immunoassays can be detectably labeled in various ways. Examples of types of immunoassays which can utilize antibodies of the invention are competitive and non-competitive immunoassays in either a direct or indirect format. Examples of such immunoassays are the radioimmunoassay (RIA) and the sandwich (immunometric) assay. Detection of the antigens using the antibodies of the invention can be done utilizing immunoassays which are run in either the forward, reverse, or simultaneous modes, including immunohistochemical assays on physiological samples. Those of skill in the art will know, or can readily discern, other immunoassay formats without undue experimentation.

The antibodies of the invention can be bound to many different carriers and used to detect the presence of an antigen comprising the polypeptide of the invention. Examples of well-known carriers include glass, polystyrene. polypropylene, polyethylene, dextran, nylon, amylases, natural and modified celluloses, polyacrylamides, agaroses and magnetite. The nature of the carrier can be either soluble or insoluble for purposes of the invention. Those skilled

in the art will know of other suitable carriers for binding antibodies, or will be able to ascertain such, using routine experimentation.

There are many different labels and methods of labeling known to those of ordinary skill in the art. Examples of the types of labels which can be used in the present invention include enzymes, radioisotopes, fluorescent compounds, colloidal metals, chemiluminescent compounds, phosphorescent compounds, and bioluminescent compounds. Those of ordinary skill in the art will know of other suitable labels for binding to the antibody, or will be able to ascertain such, using routine experimentation.

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Another technique which may also result in greater sensitivity consists of coupling the antibodies to low molecular weight haptens. These haptens can then be specifically detected by means of a second reaction. For example, it is common to use such haptens as biotin, which reacts with avidin, or dinitrophenyl, puridoxal, and fluorescein, which can react with specific antihapten antibodies.

In using the monoclonal antibodies of the invention for the *in vivo* detection of antigen, the detectably labeled antibody is given a dose which is diagnostically effective. The term "diagnostically effective" means that the amount of detectably labeled monoclonal antibody is administered in sufficient quantity to enable detection of the site having the antigen comprising a polypeptide of the invention for which the monoclonal antibodies are specific.

The concentration of detectably labeled monoclonal antibody which is adminstered should be sufficient such that the binding to those cells having the polypeptide is detectable compared to the background. Further, it is desirable that the detectably labeled monoclonal antibody be rapidly cleared from the circulatory system in order to give the best target-to-background signal ratio.

As a rule, the dosage of detectably labeled monoclonal antibody for in vivo diagnosis will vary depending on such factors as age, sex, and extent of disease of the individual. Such dosages may vary, for example, depending on whether multiple injections are given, antigenic burden, and other factors known to those of skill in the art.

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For *in vivo* diagnostic imaging, the type of detection instrument available is a major factor in selecting a given radioisotope. The radioisotope chosen must have a type of decay which is detectable for a given type of instrument. Still another important factor in selecting a radioisotope for *in vivo* diagnosis is that deleterious radiation with respect to the host is minimized. Ideally, a radioisotope used for *in vivo* imaging will lack a particle emission, but produce a large number of photons in the 140-250 keV range, which may readily be detected by conventional gamma cameras.

For *in vivo* diagnosis radioisotopes may be bound to immunoglobulin either directly or indirectly by using an intermediate functional group. Intermediate functional groups which often are used to bind radioisotopes which exist as metallic ions to immunoglobulins are the bifunctional chelating agents such as diethylenetriaminepentacetic acid (DTPA) and ethylenediaminetetraacetic acid (EDTA) and similar molecules. Typical examples of metallic ions which can be bound to the monoclonal antibodies of the invention are <sup>111</sup>In, <sup>97</sup>Ru, <sup>67</sup>Ga, <sup>68</sup>Ga, <sup>72</sup>As. <sup>89</sup>Zr, and <sup>201</sup>Tl.

The monoclonal antibodies of the invention can also be labeled with a paramagnetic isotope for purposes of *in vivo* diagnosis, as in magnetic resonance imaging (MRI) or electron spin resonance (ESR). In general, any conventional method for visualizing diagnostic imaging can be utilized. Usually gamma and positron emitting radioisotopes are used for camera imaging and

paramagnetic isotopes for MRI. Elements which are particularly useful in such techniques include <sup>157</sup>Gd, <sup>55</sup>Mn, <sup>162</sup>Dy, <sup>52</sup>Cr, and <sup>56</sup>Fe.

The monoclonal antibodies of the invention can be used *in vitro* and *in vivo* to monitor the course of amelioration of a GDF-9-associated disease in a subject. Thus, for example, by measuring the increase or decrease in the number of cells expressing antigen comprising a polypeptide of the invention or changes in the concentration of such antigen present in various body fluids, it would be possible to determine whether a particular therapeutic regimen aimed at ameliorating the GDF-9-associated disease is effective. The term "ameliorate" denotes a lessening of the detrimental effect of the GDF-9-associated disease in the subject receiving therapy.

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The present invention identifies a nucleotide sequence that can be expressed in an altered manner as compared to expression in a normal cell, therefore, it is possible to design appropriate therapeutic or diagnostic techniques directed to this sequence. Thus, where a cell-proliferative disorder is associated with the expression of GDF-9, nucleic acid sequences that interfere with GDF-9 expression at the translational level can be used. This approach utilizes, for example, antisense nucleic acid and ribozymes to block translation of a specific GDF-9 mRNA, either by masking that mRNA with an antisense nucleic acid or by cleaving it with a ribozyme.

Antisense nucleic acids are DNA or RNA molecules that are complementary to at least a portion of a specific mRNA molecule (Weintraub, *Scientific American*, 262:40, 1990). In the cell, the antisense nucleic acids hybridize to the corresponding mRNA, forming a double-stranded molecule. The antisense nucleic acids interfere with the translation of the mRNA, since the cell will not translate a mRNA that is double-stranded. Antisense oligomers of about 15 nucleotides are preferred, since they are easily synthesized and are less likely

to cause problems than larger molecules when introduced into the target GDF-The use of antisense methods to inhibit the in vitro 9-producing cell. translation of genes is well known in the art (Marcus-Sakura, Anal.Biochem., 172:289, 1988).

Ribozymes are RNA molecules possessing the ability to specifically cleave other single-stranded RNA in a manner analogous to DNA restriction Through the modification of nucleotide sequences which endonucleases. encode these RNAs, it is possible to engineer molecules that recognize specific nucleotide sequences in an RNA molecule and cleave it (Cech, J.Amer.Med. Assn., 260:3030, 1988). A major advantage of this approach is that, because 10 they are sequence-specific, only mRNAs with particular sequences are inactivated.

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There are two basic types of ribozymes namely, tetrahymena-type (Hasselhoff, Nature, 334:585, 1988) and "hammerhead"-type. Tetrahymena-type ribozymes recognize sequences which are four bases in length, while "hammerhead"-type ribozymes recognize base sequences 11-18 bases in length. The longer the recognition sequence, the greater the likelihood that the sequence will occur exclusively in the target mRNA species. Consequently, hammerhead-type ribozymes are preferable to tetrahymena-type ribozymes for inactivating a specific mRNA species and 18-based recognition sequences are preferable to shorter recognition sequences.

The present invention also provides gene therapy for the treatment of cell proliferative disorders which are mediated by GDF-9 protein. Such therapy would achieve its therapeutic effect by introduction of the GDF-9 antisense polynucleotide into cells having the proliferative disorder. Delivery of antisense GDF-9 polynucleotide can be achieved using a recombinant expression vector such as a chimeric virus or a colloidal dispersion system.

Especially preferred for therapeutic delivery of antisense sequences is the use of targeted liposomes.

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Various viral vectors which can be utilized for gene therapy as taught herein include adenovirus, herpes virus, vaccinia, or, preferably, an RNA virus such as a retrovirus. Preferably, the retroviral vector is a derivative of a murine or avian retrovirus. Examples of retroviral vectors in which a single foreign gene can be inserted include, but are not limited to: Moloney murine leukemia virus (MoMuLV), Harvey murine sarcoma virus (HaMuSV), murine mammary tumor virus (MuMTV), and Rous Sarcoma Virus (RSV). A number of additional retroviral vectors can incorporate multiple genes. All of these vectors can transfer or incorporate a gene for a selectable marker so that transduced cells can be identified and generated. By inserting a GDF-9 sequence of interest into the viral vector, along with another gene which encodes the ligand for a receptor on a specific target cell, for example, the vector is now target specific. Retroviral vectors can be made target specific by inserting, for example, a polynucleotide encoding a sugar, a glycolipid, or a protein. Preferred targeting is accomplished by using an antibody to target the retroviral vector. Those of skill in the art will know of, or can readily ascertain without undue experimentation, specific polynucleotide sequences which can be inserted into the retroviral genome to allow target specific delivery of the retroviral vector containing the GDF-9 antisense polynucleotide.

Since recombinant retroviruses are defective, they require assistance in order to produce infectious vector particles. This assistance can be provided, for example, by using helper cell lines that contain plasmids encoding all of the structural genes of the retrovirus under the control of regulatory sequences within the LTR. These plasmids are missing a nucleotide sequence which enables the packaging mechanism to recognize an RNA transcript for encapsidation. Helper cell lines which have deletions of the packaging signal

include, but are not limited to  $\Psi$ 2, PA317 and PA12, for example. These cell lines produce empty virions, since no genome is packaged. If a retroviral vector is introduced into such cells in which the packaging signal is intact, but the structural genes are replaced by other genes of interest, the vector can be packaged and vector virion produced.

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Alternatively, NIH 3T3 or other tissue culture cells can be directly transfected with plasmids encoding the retroviral structural genes *gag*, *pol* and *env*, by conventional calcium phosphate transfection. These cells are then transfected with the vector plasmid containing the genes of interest. The resulting cells release the retroviral vector into the culture medium.

Another targeted delivery system for GDF-9 antisense polynucleotides is a colloidal dispersion system. Colloidal dispersion systems include macromolecule complexes, nanocapsules, microspheres, beads, and lipid-based systems including oil-in-water emulsions, micelles, mixed micelles, and liposomes. The preferred colloidal system of this invention is a liposome. Liposomes are artificial membrane vesicles which are useful as delivery vehicles in vitro and in vivo. It has been shown that large unilamellar vesicles (LUV), which range in size from 0.2-4.0  $\mu m$  can encapsulate a substantial percentage of an aqueous buffer containing large macromolecules. RNA, DNA and intact virions can be encapsulated within the aqueous interior and be delivered to cells in a biologically active form (Fraley, et al., Trends Biochem. Sci., 6:77, 1981). In addition to mammalian cells, liposomes have been used for delivery of polynucleotides in plant, yeast and bacterial cells. In order for a liposome to be an efficient gene transfer vehicle, the following characteristics should be present: (1) encapsulation of the genes of interest at high efficiency while not compromising their biological activity; (2) preferential and substantial binding to a target cell in comparison to non-target cells; (3) delivery of the aqueous contents of the vesicle to the target cell cytoplasm at high efficiency; and (4) accurate and effective expression of genetic information (Mannino, et al., Biotechniques, 6:682, 1988).

The composition of the liposome is usually a combination of phospholipids, particularly high-phase-transition-temperature phospholipids, usually in combination with steroids, especially cholesterol. Other phospholipids or other lipids may also be used. The physical characteristics of liposomes depend on pH, ionic strength, and the presence of divalent cations.

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Examples of lipids useful in liposome production include phosphatidyl compounds, such as phosphatidylglycerol, phosphatidylcholine, phosphatidylserine, phosphatidylethanolamine, sphingolipids, cerebrosides, and gangliosides. Particularly useful are diacylphosphatidylglycerols, where the lipid moiety contains from 14-18 carbon atoms, particularly from 16-18 carbon atoms, and is saturated. Illustrative phospholipids include egg phosphatidylcholine, dipalmitoylphosphatidylcholine and distearoylphosphatidylcholine.

The targeting of liposomes can be classified based on anatomical and mechanistic factors. Anatomical classification is based on the level of selectivity, for example, organ-specific, cell-specific, and organelle-specific. Mechanistic targeting can be distinguished based upon whether it is passive or active. Passive targeting utilizes the natural tendency of liposomes to distribute to cells of the reticulo-endothelial system (RES) in organs which contain sinusoidal capillaries. Active targeting, on the other hand, involves alteration of the liposome by coupling the liposome to a specific ligand such as a monoclonal antibody, sugar, glycolipid, or protein, or by changing the composition or size of the liposome in order to achieve targeting to organs and cell types other than the naturally occurring sites of localization.

The surface of the targeted delivery system may be modified in a variety of ways. In the case of a liposomal targeted delivery system, lipid groups can be incorporated into the lipid bilayer of the liposome in order to maintain the targeting ligand in stable association with the liposomal bilayer. Various linking groups can be used for joining the lipid chains to the targeting ligand.

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Due to the expression of GDF-9 in the reproductive tract, there are a variety of applications using the polypeptide, polynucleotide and antibodies of the invention, related to contraception, fertility and pregnancy. GDF-9 could play a role in regulation of the menstrual cycle and, therefore, could be useful in various contraceptive regimens.

The following examples are intended to illustrate but not limit the invention. While they are typical of those that might be used, other procedures known to those skilled in the art may alternatively be used.

# EXAMPLE 1 IDENTIFICATION AND ISOLATION OF A NOVEL TGF-B FAMILY MEMBER

To identify a new member of the TGF- $\beta$  superfamily, degenerate oligonucleotides were designed which corresponded to two conserved regions among the known family members: one region spanning the two tryptophan residues conserved in all family members except MIS and the other region spanning the invariant cysteine residues near the C-terminus. These primers were used for polymerase chain reactions on mouse genomic DNA followed by subcloning the PCR products using restriction sites placed at the 5' ends of the primers, picking individual *E. coli* colonies carrying these subcloned inserts, and using a combination of random sequencing and hybridization analysis to eliminate known members of the superfamily.

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GDF-9 was identified from a mixture of PCR products obtained with the primers SJL160 (5'-CCGGAATTCGGITGG(G/C/A)A(G/A/T/C)(G/C/A)A(G/A/T/C) TGG(A/G)TI(T/G)CICC-3') (SEQUENCE ID NO. 1) and SJL153 (5'-C C G G A A T T C ( A / G ) C A I ( G / C ) C ( A / G ) C A I C ( T / C ) ( G / A / T - /C)(C/G/T)TIG(T/C)I(G/A)(T/C)CAT-3') (SEQUENCE ID NO. 2). PCR using these primers was carried out with 2  $\mu$ g mouse genomic DNA at 94°C for 1 min, 50°C for 2 min, and 72°C for 2 min for 40 cycles.

PCR products of approximately 280 bp were gel-purified, digested with Eco RI, gel-purified again, and subcloned in the Bluescript vector (Stratagene, San Diego, CA). Bacterial colonies carrying individual subclones were picked into 96 well microtiter plates, and multiple replicas were prepared by plating the cells onto nitrocellulose. The replicate filters were hybridized to probes representing known members of the family, and DNA was prepared from non-hybridizing colonies for sequence analysis.

The primer combination of SJL160 and SJL153, yielded three known sequences (inhibin  $\beta$ B, BMP-2, and BMP-4) and one novel sequence (designated GDF-9) among 145 subclones analyzed.

RNA isolation and Northern analysis were carried out as described previously (Lee,S.J., *Mol. Endocrinol.* 4:1034, 1990). An oligo dT-primed cDNA library was prepared from 2.5-3  $\mu$ g of ovary poly A-selected RNA in the lambda ZAP II vector according to the instructions provided by Stratagene. The ovary library was not amplified prior to screening. Filters were hybridized as described previously (Lee, S.-J., *Proc. Natl. Acad. Sci. USA.*, 88:4250-4254, 1991). DNA sequencing of both strands was carried out using the dideoxy chain termination method (Sanger, *et al., Proc. Natl. Acad. Sci., USA*, 74:5463-5467, 1977) and a combination of the S1 nuclease/exonuclease III strategy (Henikoff, S., *Gene*, 28:351-359, 1984) and synthetic oligonucleotide primers.

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## EXAMPLE 2 EXPRESSION PATTERN AND SEQUENCE OF GDF-9

To determine the expression pattern of GDF-9, RNA samples prepared from a variety of adult tissues were screened by Northern analysis. Five micrograms of twice polyA-selected RNA prepared from each tissue were electrophoresed on formaldehyde gels, blotted and probed with GDF-9. As shown in Figure 1, the GDF-9 probe detected a 1.7 kb mRNA expressed exclusively in the ovary.

A mouse ovary cDNA library of  $1.5 \times 10^6$  recombinant phage was constructed in lambda ZAP II and screened with a probe derived from the GDF-9 PCR product. The nucleotide sequence of the longest of nineteen hybridizing clones is shown in Figure 2. Consensus N-glycosylation signals are denoted by plain boxes. The putative tetrabasic processing sites are denoted by

stippled boxes. The in-frame termination codons upstream of the putative initiating ATG and the consensus polyadenylation signals are underlined. The poly A tails are not shown. Numbers indicate nucleotide position relative to the 5' end. The 1712 bp sequence contains a long open reading frame beginning with a methionine codon at nucleotide 29 and potentially encoding a protein 441 amino acids in length with a molecular weight of 49.6 kD. Like other TGF- $\beta$  family members, the GDF-9 sequence contains a core of hydrophobic amino acids near the N-terminus suggestive of a signal sequence for secretion. GDF-9 contains four potential N-glycosylation sites at asparagine residues 163, 229, 258, and 325 and a putative tetrabasic proteolytic processing site (RRRR) at amino acids 303-306. The mature C-terminal fragment of GDF-9 is predicted to be 135 amino acids in length and have an unglycosylated molecular weight of 15.6 kD.

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Although the C-terminal portion of GDF-9 clearly shows homology with the other family members, the sequence of GDF-9 is significantly diverged from those of the other family members (Figures 3 and 4). Figure 3 shows the alignment of the C-terminal sequences of GDF-9 with the corresponding regions of human GDF-1 (Lee, Proc. Natl. Acad. Sci. USA, 88:4250-4254, 1991), Xenopus Vg-1 (Weeks, et al., Cell, 51:861-867, 1987), human Vgr-1 (Celeste, et al., Proc. Natl. Acad. Sci. USA, 87:9843-9847, 1990), human OP-1 (Ozkaynak, et al., EMBO J., 9:2085-2093, 1990), human BMP-5 (Celeste, et al., Proc. Natl. Acad. Sci. USA, 87:9843-9847, 1990), Drosophila 60A (Wharton, et al., Proc. Natl. Acad. Sci. USA, 88:9214-9218, 1991), human BMP-2 and 4 (Wozney, et al., Science, 242:1528-1534, 1988), Drosophila DPP (Padgett, et al., Nature, 325:81-84, 1987), human BMP-3 (Wozney, et al., Science, 242:1528-1534, 1988), human MIS (Cate, et al., Cell, 45:685-698, 1986), human inhibin,  $\beta A$ , and  $\beta B$  (Mason, et al., Biochem, Biophys. Res. Commun., 135:957-964, 1986), human TGF-β1 (Derynck, et al., Nature, <u>316</u>:701-705, 1985), humanTGF-β2 (deMartin, et al., EMBO J., 6:3673-3677, 1987), human TGF-β3

(ten Dijke, et al., Proc. Natl. Acad. Sci. USA, <u>85</u>:4715-4719, 1988), chicken TGFβ4 (Jakowlew, et al., Mol. Endocrinol., <u>2</u>:1186-1195, 1988), and Xenopus TGFβ5 (Kondaiah, et al., J. Biol. Chem., <u>265</u>:1089-1093, 1990). The conserved cysteine residues are shaded. Dashes denote gaps introduced in order to maximize the alignment.

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Figure 4 shows the amino acid homologies among the different members of the TGF- $\beta$  superfamily. Numbers represent percent amino acid identities between each pair calculated from the first conserved cysteine to the C-terminus. Boxes represent homologies among highly-related members within particular subgroups.

The degree of sequence identify with known family members ranges from a minimum of 21% with MIS to a maximum of 34% with BMP-4. Hence, GDF-9 is comparable to MIS in its degree of sequence divergence from the other members of this superfamily. Moreover, GDF-9 shows no significant sequence homology to other family members in the pro-region of the molecule. GDF-9 also differs from the known family members in its pattern of cysteine residues in the C-terminal region. GDF-9 lacks the fourth cysteine of the seven cysteines that are present in all other family members; in place of cysteine at this position, the GDF-9 sequence contains a serine residue. In addition, GDF-9 does not contain a seventh cysteine residue elsewhere in the C-terminal region.

# EXAMPLE 3 IMMUNOCHEMICAL LOCALIZATION OF GDF-9 IN THE ZONA PELLUCIDA

To determine whether GDF-9 mRNA was translated, sections of adult ovaries were incubated with antibodies directed against recombinant GDF-9 protein. In order to raise antibodies against GDF-9, portions of GDF-9 cDNA spanning amino acids 30 to 295 (pro-region) or 308 to 441 (mature region) were cloned into the T7-based pET3 expression vector (provided by F.W. Studier, Brookhaven National Laboratory), and the resulting plasmids were transformed into the BL21 (DE3) bacterial strain. Total cell extracts from isopropyl B-D-thiogalactoside-induced cells were electrophoresed on SDS/polyacrylamide gels, and the GDF-9 protein fragments were excised, mixed with Freund's adjuvant, and used to immunize rabbits by standard methods known to those of skill in the art. All immunizations were carried out by Spring Valley Lab (Sykesville, MD). The presence of GDF-9-reactive antibodies in the sera of these rabbits was assessed by Western analysis of bacterially-expressed protein fragments. The resulting serum was shown to react with the bacterially-expressed protein by Western analysis.

For immunohistochemical studies, ovaries were removed from adult mice, fixed in 4% paraformaldehyde, embedded in paraffin, and sectioned. Sites of antibody binding were detected by using the Vectastain ABC kit, according to the instructions provided by Vector Laboratories. FIGURE 5 shows the immunohistochemical localization of GDF-9 protein. Adjacent sections of an adult ovary were either stained with hematoxylin and eosin (FIGURE 5a) or incubated with immune (FIGURE 5b) or pre-immune (FIGURE 5c) serum at a dilution of 1:500. As shown in FIGURE 5b, the antiserum detected protein

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solely in oocytes. No staining was detected using pre-immune serum (FIGURE 5c). Hence, GDF-9 protein appears to translated *in vivo* by oocytes.

# EXAMPLE 4 ISOLATION OF HUMAN GDF-9

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In order to isolate a cDNA clone encoding human GDF-9, a cDNA library was constructed in lambda ZAP II using poly A-selected RNA prepared from an adult human ovary. From this library, a cDNA clone containing the entire human GDF-9 coding sequence was identified using standard screening techniques as in Example 1 and using the murine GDF-9 clone as a probe. A comparison of the predicted amino acid sequences of murine (top lines) and human (bottom lines) GDF-9 is shown in FIGURE 6. Numbers represent amino acid positions relative to the N-termini. Vertical lines represent sequence identities. Dots represent gaps introduced in order to maximize the alignment. The clear box shows the predicted proteolytic processing sites. The shaded boxes show the cysteine residues in the mature region of the proteins. The bars at the bottom show a schematic of the pre-(clear) and mature (shaded) regions of GDF-9 with the percent sequence identities between the murine and human sequences shown below.

Like murine GDF-9, human GDF-9 contains a hydrophobic leader sequence, a putative RXXR proteolytic cleavage site, and a C-terminal region containing the hallmarks of other TGF- $\beta$  family members. Murine and human GDF-9 are 64% identical in the pro- region and 90% identical in the predicted mature region of the molecule. The high degree of homology between the two sequences suggests that human GDF-9 plays an important role during embryonic development and/or in the adult ovary.

#### **EXAMPLE 5**

#### NUCLEIC ACID DETECTION OF EXPRESSION OF GDF-9 IN OOCYTES

In order to localize the expression of GDF-9 in the ovary, *in situ* hybridization to mouse ovary sections was carried out using an antisense GDF-9 RNA probe. FIGURE 7 shows *in situ* hybridization to adult ovary sections using a GDF-9 RNA probe. [<sup>35</sup>S]-labeled anti-sense (FIGURE 7a and 7c) or sense (FIGURE 7 b and 7d) GDF-9 RNA probes were hybridized to adjacent paraffinembedded sections of ovaries fixed in 4% paraformaldehyde. Sections were dipped in photographic emulsion, exposed, developed, and then stained with hematoxylin and eosin. Two representative fields are shown.

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As shown in FIGURES 7a and 7c, GDF-9 mRNA was detected primarily in oocytes in adult ovaries. Every oocyte (regardless of the stage of follicular development) examined showed GDF-9 expression, and no expression was detected in any other cell types. No hybridization was seen using a control GDF-9 sense RNA probe (FIGURE 7b and 7d). Hence, GDF-9 expression appears to be oocyte-specific in adult ovaries.

To determine the pattern of expression of GDF-9 mRNA during ovarian development, sections of neonatal ovaries were probed with a GDF-9 RNA probe. FIGURE 8 shows *in situ* hybridization to a postnatal day 4 ovary section using an antisense GDF-9 RNA probe. Sections were prepared as described for FIGURE 7. Following autoradiography and staining, the section was photographed under bright-field (FIGURE 8a) or dark-field (FIGURE 8b) illumination.

FIGURE 9 shows *in situ* hybridization to postnatal day 8 ovary sections using an antisense (FIGURE 9a) or sense (FIGURE 9b) GDF-9 RNA probe. Sections were prepared as described for FIGURE 7.

GDF-9 mRNA expression was first detected at the onset of follicular development. This was most clearly evident at postnatal day 4, where only occytes that were present in follicles showed GDF-9 expression (FIGURE 8); no expression was seen in occytes that were not surrounded by granulosa cells. By postnatal day 8, every occyte appeared to have undergone follicular development, and every occyte showed GDF-9 expression (FIGURE 9).

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To determine whether GDF-9 was also expressed following ovulation, sections of mouse oviducts were examined by *in situ* hybridization. FIGURE 10 shows *in situ* hybridization to adult oviduct sections using an antisense (FIGURE 10a) or sense (FIGURE 10b) GDF-9 RNA probe. Sections were prepared as described for FIGURE 7.

FIGURE 11 shows *in situ* hybridization to an adult oviduct (0.5 days following fertilization) section using an antisense GDF-9 RNA probe. Sections were prepared as described for FIGURE 7. Following autoradiography and staining, the section was photographed under bright-field (FIGURE 11a) or dark-field (FIGURE 11b) illumination.

As shown in FIGURE 10, GDF-9 was expressed by oocytes that had been released into the oviduct. However, the expression of GDF-9 mRNA turned off rapidly following fertilization of the oocytes; by day 0.5 following fertilization, only some embryos (such as the one shown in FIGURE 11) expressed GDF-9 mRNA, and by day 1.5, all embryos were negative for GDF-9 expression.

Although the invention has been described with reference to the presently preferred embodiment, it should be understood that various modifications can be made without departing from the spirit of the invention. Accordingly, the invention is limited only by the following claims.

#### SUMMARY OF SEQUENCES

Sequence ID No. 1 is the nucleotide sequence for the primer, SJL160, for GDF-9 (page 24, lines 15 and 16);

Sequence ID No. 2 is the nucleotide sequence for the primer, SJL153, for GDF-9 (page 24, lines 17 and 18);

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Sequence ID No. 3 is the nucleotide and deduced amino acid sequence for GDF-9 (Figure 2);

Sequence ID No. 4 is the deduced amino acid sequence for GDF-9 (Figure 2);

Sequence ID No. 5 is the amino acid sequence of the C-terminus of GDF-3 (Figure 3);

Sequence ID No. 6 is the amino acid sequence of the C-terminus of GDF-9 (Figure 3);

Sequence ID No. 7 is the amino acid sequence of the C-terminus of GDF-1 (Figure 3);

Sequence ID No. 8 is the amino acid sequence of the C-terminus of Vg-1 (Figure 3);

Sequence ID No. 9 is the amino acid sequence of the C-terminus of Vgr-1 (Figure 3);

Sequence ID No. 10 is the amino acid sequence of the C-terminus of OP-1 (Figure 3);

Sequence ID No. 11 is the amino acid sequence of the C-terminus of BMP-5 (Figure 3);

Sequence ID No. 12 is the amino acid sequence of the C-terminus of 60A (Figure 3);

Sequence ID No. 13 is the amino acid sequence of the C-terminus of BMP-2 (Figure 3);

Sequence ID No. 14 is the amino acid sequence of the C-terminus of BMP-4 (Figure 3);

Sequence ID No. 15 is the amino acid sequence of the C-terminus of DPP (Figure 3);

Sequence ID No. 16 is the amino acid sequence of the C-terminus of BMP-3 (Figure 3);

Sequence ID No. 17 is the amino acid sequence of the C-terminus of MIS (Figure 3);

Sequence ID No. 18 is the amino acid sequence of the C-terminus of inhibin  $\alpha$  (Figure 3);

Sequence ID No. 19 is the amino acid sequence of the C-terminus of inhibin  $\beta$ A (Figure 3);

Sequence ID No. 20 is the amino acid sequence of the C-terminus of inhibin 20 \$\beta\$B (Figure 3); Sequence ID No. 21 is the amino acid sequence of the C-terminus of TGF- $\beta$ 1 (Figure 3);

Sequence ID No. 22 is the amino acid sequence of the C-terminus of TGF- $\beta$ 2 (Figure 3);

5 Sequence ID No. 23 is the amino acid sequence of the C-terminus of TGF-β3 (Figure 3);

Sequence ID No. 24 is the amino acid sequence of the C-terminus of TGF- $\beta$ 4 (Figure 3);

Sequence ID No. 25 is the amino acid sequence of the C-terminus of TGF- $\beta$ 5 (Figure 3); and

Sequence ID No. 26 is the amino acid sequence of human GDF-9 (Figure 6).

## SEQUENCE LISTING

	(1) GENERAL INFORMATION:	
	(1) APPLICANT: THE JOHNS HO	PKINS UNIVERSITY
	(ii) TITLE OF INVENTION: GRO	WTH DIFFERENTIATION FACTOR-9
5	(iii) NUMBER OF SEQUENCES: 26	
	(iv) CORRESPONDENCE ADDRESS:	
	(A) ADDRESSEE: Spensle	y Horn Jubas & Lubitz
	(B) STREET: 1880 Centu	ry Park East, Suite 500
10	(C) CITY: Los Angeles	
	(D) STATE: California	
	(E) COUNTRY: US	
	(F) ZIP: 90067	
	(v) COMPUTER READABLE FORM:	
15	(A) MEDIUM TYPE: Flopp	y disk
	(B) COMPUTER: IBM PC c	ompatible
	(C) OPERATING SYSTEM:	_ ·
	(D) SOFTWARE: PatentIn	Release #1.0, Version #1.25
	(vi) CURRENT APPLICATION DAT	A:
20	(A) APPLICATION NUMBER	
	(B) FILING DATE: 12-JA	N-1994
	(C) CLASSIFICATION:	
	(viii) ATTORNEY/AGENT INFORMAT	
	(A) NAME: Wetherell, J	
25	(B) REGISTRATION NUMBE	
	(C) REFERENCE/DOCKET N	UMBER: FD3288
	(ix) TELECOMMUNICATION INFOR	MATION:

## 30 (2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 35 base pairs

(A) TELEPHONE: (619) 455-5100 (B) TELEFAX: (619) 455-5110

(B) TYPE: nucleic acid

```
(C) STRANDEDNESS: single
                 (D) TOPOLOGY: linear
           (ii) MOLECULE TYPE: DNA (genomic)
          (vii) IMMEDIATE SOURCE:
                 (B) CLONE: SJL160
5
           (ix) FEATURE:
                 (A) NAME/KEY: CDS
                 (B) LOCATION: 1..35
                 (D) OTHER INFORMATION: /note= "Where "B" occurs, B =
10
                        inosine"
           (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
                                                                                35
       CCGGAATTCG GBTGGVANVA NTGGRTBRTB KCBCC
       (2) INFORMATION FOR SEQ ID NO:2:
            (i) SEQUENCE CHARACTERISTICS:
                 (A) LENGTH: 33 base pairs
15
                 (B) TYPE: nucleic acid
                 (C) STRANDEDNESS: single
                 (D) TOPOLOGY: linear
           (ii) MOLECULE TYPE: DNA (genomic)
20
          (vii) IMMEDIATE SOURCE:
                 (B) CLONE: SJL153
           (ix) FEATURE:
                 (A) NAME/KEY: CDS
                 (B) LOCATION: 1..33
25
           (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
                                                                                33
       CCGGAATTCR CADSCRCADC YNBTDGYDRY CAT
       (2) INFORMATION FOR SEQ ID NO:3:
```

(i) SEQUENCE CHARACTERISTICS:

5	(A) LENGTH: 1712 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear  (ii) MOLECULE TYPE: DNA (genomic)	·
	(vii) IMMEDIATE SOURCE: (B) CLONE: GDF-9	
10	(ix) FEATURE:  (A) NAME/KEY: CDS  (B) LOCATION: 291351	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:	
15	ATGCGTTCCT TCTTAGTTCT TCCAAGTC ATG GCA CTT CCC AGC AAC TTC CTG  Met Ala Leu Pro Ser Asn Phe Leu  1 5	52
	TTG GGG GTT TGC TGC TTT GCC TGG CTG TGT TTT CTT AGT AGC CTT AGC Leu Gly Val Cys Cys Phe Ala Trp Leu Cys Phe Leu Ser Ser Leu Ser 10 15 20	100
20	TCT CAG GCT TCT ACT GAA GAA TCC CAG AGT GGA GCC AGT GAA AAT GTG Ser Gln Ala Ser Thr Glu Glu Ser Gln Ser Gly Ala Ser Glu Asn Val 25 30 35 40	148
	GAG TCT GAG GCA GAC CCC TGG TCC TTG CTG CTG CCT GTA GAT GGG ACT Glu Ser Glu Ala Asp Pro Trp Ser Leu Leu Pro Val Asp Gly Thr 45 50 55	196
25	GAC AGG TCT GGC CTC TTG CCC CCC CTC TTT AAG GTT CTA TCT GAT AGG Asp Arg Ser Gly Leu Leu Pro Pro Leu Phe Lys Val Leu Ser Asp Arg 60 65 70	244
30	CGA GGT GAG ACC CCT AAG CTG CAG CCT GAC TCC AGA GCA CTC TAC TAC Arg Gly Glu Thr Pro Lys Leu Gln Pro Asp Ser Arg Ala Leu Tyr Tyr 75 80 85	292

										GAG Glu 100					340	
5										CTC Leu					388	,
										ACA Thr					436	1
10										ACT Thr					<b>4</b> 84	•
15										AAC Asn					532	)
										GTA Val 180					580	)
20										TCA Ser					628	3
									Ser	CTC Leu				CTA Leu	676	5
25				Glu				Leu		GTC Val			Thr		724	4
30			Gln				Gly					Pro		TCA Ser	77:	2
		Pro				Tyr					Ser			GCC Ala	829	0

	TAC Tyr 265									TGG Trp 275						868
5	CCC Pro									GTG Val						916
										GGG Gly						964
10										TCC Ser						1012
15										GAG Glu						1060
										GAC Asp 355						1108
20										GGG Gly						1156
					Tyr				His					Asn	ATA Ile	1204
25				Lys				Val					Cys		CCG Pro	1252
30			Tyr				Val					Pro			TCC Ser	1300
		Ala				Glu					Thr				TGT Cys 440	1348

	CGT TAGCATGGGG GCCACTTCAA CAAGCCTGCC TGGCAGAGCA ATGCTGTGGG Arg	1401
	CCTTAGAGTG CCTGGGCAGA GAGCTTCCTG TGACCAGTCT CTCCGTGCTG CTCAGTGCAC	1461
5	ACTGTGTGAG CGGGGGAAGT GTGTGTGTT GGATGAGCAC ATCGAGTGCA GTGTCCGTAG	1521
	GTGTAAAGGG CACACTCACT GGTCGTTGCC ATAAACCAAG TGAAATGTAA CTCATTTGGA	1581
	GAGCTCTTTC TCCCCACGAG TGTAGTTTTC AGTGGACAGA TTTGTTAGCA TAAGTCTCGA	1641
	GTAGAATGTA GCTGTGAACA TGTCAGAGTG CTGTGGTTTT ATGTGACGGA AGAATAAACT	1701
	GTTGATGGCA T	1712
10	(2) INFORMATION FOR SEQ ID NO:4:  (i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 441 amino acids  (B) TYPE: amino acid  (D) TOPOLOGY: linear	
15	(ii) MOLECULE TYPE: protein	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
	Met Ala Leu Pro Ser Asn Phe Leu Leu Gly Val Cys Cys Phe Ala Trp  1 5 10 15	
20	Leu Cys Phe Leu Ser Ser Leu Ser Ser Gln Ala Ser Thr Glu Glu Ser 20 25 30	
	Gln Ser Gly Ala Ser Glu Asn Val Glu Ser Glu Ala Asp Pro Trp Ser 35 40 45	
	Leu Leu Pro Val Asp Gly Thr Asp Arg Ser Gly Leu Leu Pro Pro 50 55 60	
25	Leu Phe Lys Val Leu Ser Asp Arg Gly Glu Thr Pro Lys Leu Gln 65 70 75 80	
	Pro Asp Ser Arg Ala Leu Tyr Tyr Met Lys Lys Leu Tyr Lys Thr Tyr 85 90 95	

	Ala	inr	Lys	100	Gly	vai	110	Lys	105	Jei	ur 6	DCI		110	-,-	
	Thr	Val	Arg 115	Leu	Phe	Ser	Pro	Cys 120	Ala	Gln	Gln	Glu	Gln 125	Ala	Pro	Ser
5	Asn	Gln 130	Val	Thr	Gly	Pro	Leu 135	Pro	Met	Val	Asp	Leu 140	Leu	Phe	Asn	Leu
	Asp 145	Arg	Val	Thr	Ala	Met 150	Glu	His	Leu	Leu	Lys 155	Ser	Val	Leu	Leu	Tyr 160
10	Thr	Leu	Asn	Asn	Ser 165	Ala	Ser	Ser	Ser	Ser 170	Thr	Val	Thr	Cys	Met 175	Cys
	Asp	Leu	Val	Val 180	Lys	Glu	Ala	Met	Ser 185	Ser	Gly	Arg	Ala	Pro 190	Pro	Arg
	Ala	Pro	Tyr 195	Ser	Phe	Thr	Leu	Lys 200	Lys	His	Arg	Trp	11e 205	Glu	Ile	Asp
15	Val	Thr 210	Ser	Leu	Leu	Gln	Pro 215	Leu	Val	Thr	Ser	Ser 220	Glu	Arg	Ser	Ile
	His 225	Leu	Ser	Val	Asn	Phe 230	Thr	Cys	Thr	Lys	Asp 235	Gln	Val	Pro	Glu	Asp 240
20	Gly	Val	Phe	Ser	Met 245	Pro	Leu	Ser	Val	Pro 250	Pro	Ser	Leu	Ile	Leu 255	Tyr
	Leu	Asn	Asp	Thr 260	Ser	Thr	Gln	Ala	Tyr 265	His	Ser	Trp	Gln	Ser 270	Leu	Gln
	Ser	Thr	Trp 275	_	Pro	Leu	Gln	His 280		Gly	Gln	Ala	Gly 285	Val	Ala	Ala
25	Arg	Pro 290		Lys	Glu	Glu	Ala 295		Glu	Val	Glu	Arg 300		Pro	Arg	Arg
	Arg 305	_	Gly	Gln	Lys	Ala 310		Arg	Ser	Glu	Ala 315	Lys	Gly	Pro	Leu	120 320
30	Thr	Ala	Ser	Phe	Asn		Ser	Glu	Tyr	Phe 330		Gln	Phe	Leu	Phe	

	Gln	Asn	Glu	Cys 340	Glu	Leu	His	Asp	Phe 345	Arg	Leu	Ser	Phe	Ser 350	Gln	Leu	
	Lys	Trp	Asp 355	Asn	Trp	Ile	Val	Ala 360	Pro	His	Arg	Tyr	Asn 365	Pro	Arg	Tyr	
5	Cys	Lys 370	Gly	Asp	Cys	Pro	Arg 375	Ala	Val	Arg	His	Arg 380	Tyr	Gly	Ser	Pro	
	Val 385	His	Thr	Met	Val	Gln 390	Asn	Ile	Ile	Tyr	Glu 395	Lys	Leu	Asp	Pro	Ser 400	
10	Val	Pro	Arg	Pro	Ser 405	Cys	Val	Pro	Gly	Lys 410	Tyr	Ser	Pro	Leu	Ser 415	Val	
	Leu	Thr	Ile	Glu 420	Pro	Asp	Gly	Ser	Ile 425	Ala	Tyr	Lys	Glu	Tyr 430	Glu	Asp .	
	Met	Ile	Ala 435	Thr	Arg	Cys	Thr	Cys 440	Arg								
15	(2)	INF	ORMA'	TION	FOR	SEQ	ID	NO:5	:								
20		·	(; (; (;	A) L B) T C) S D) T	CE CI ENGTI YPE: TRANI OPOLO	H: 1: amin DEDN: OGY:	17 am no a ESS: lin	mino cid sin ear	aci	ds							
		(vii			ATE :												
<b>2</b> 5		(ix		A) N	E: AME/I												
		(xi	) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:5:						
		Ly 1	s Ar	g Ar	g Al	a Al 5	a Il	e Se	r Va	l Pr	o Ly 10		y Ph	е Су	s Ar	g Asn 15	Phe

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Cys His Arg His Gln Leu Phe Ile Asn Phe Gln Asp Leu Gly Trp His 25 20 Lys Trp Val Ile Ala Pro Lys Gly Phe Met Ala Asn Tyr Cys His Gly 45 40 35 Glu Cys Pro Phe Ser Met Thr Thr Tyr Leu Asn Ser Ser Asn Tyr Ala 5 60 55 50 Phe Met Gln Ala Leu Met His Met Ala Asp Pro Lys Val Pro Lys Ala 75 70 65 Val Cys Val Pro Thr Lys Leu Ser Pro Ile Ser Met Leu Tyr Gln Asp 95 90 85 10 Ser Asp Lys Asn Val Ile Leu Arg His Tyr Glu Asp Met Val Val Asp 110 105 100 Glu Cys Gly Cys Gly 115 (2) INFORMATION FOR SEQ ID NO:6: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: GDF-9 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..118 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6: Phe Asn Leu Ser Glu Tyr Phe Lys Gln Phe Leu Phe Pro Gln Asn Glu 15 10

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Cys Glu Leu His Asp Phe Arg Leu Ser Phe Ser Gln Leu Lys Trp Asp 25 20 Asn Trp Ile Val Ala Pro His Arg Tyr Asn Pro Arg Tyr Cys Lys Gly 45 35 40 Asp Cys Pro Arg Ala Val Arg His Arg Tyr Gly Ser Pro Val His Thr 5 60 50 55 Met Val Gln Asn Ile Ile Tyr Glu Lys Leu Asp Pro Ser Val Pro Arg 75 70 65 Pro Ser Cys Val Pro Gly Lys Tyr Ser Pro Leu Ser Val Leu Thr Ile 90 95 85 10 Glu Pro Asp Gly Ser Ile Ala Tyr Lys Glu Tyr Glu Asp Met Ile Ala 105 110 100 Thr Arg Cys Thr Cys Arg 115 15 (2) INFORMATION FOR SEQ ID NO:7: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 122 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single 20 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: GDF-1 (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..122 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7: Pro Arg Arg Asp Ala Glu Pro Val Leu Gly Gly Gly Pro Gly Gly Ala

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Cys Arg Ala Arg Arg Leu Tyr Val Ser Phe Arg Glu Val Gly Trp His 25 20 Arg Trp Val Ile Ala Pro Arg Gly Phe Leu Ala Asn Tyr Cys Gln Gly 40 35 Gln Cys Ala Leu Pro Val Ala Leu Ser Gly Ser Gly Pro Pro Ala 5 55 50 Leu Asn His Ala Val Leu Arg Ala Leu Met His Ala Ala Ala Pro Gly 75 70 65 Ala Ala Asp Leu Pro Cys Cys Val Pro Ala Arg Leu Ser Pro Ile Ser 90 85 10 Val Leu Phe Phe Asp Asn Ser Asp Asn Val Val Leu Arg Gln Tyr Glu 110 105 100 Asp Met Val Val Asp Glu Cys Gly Cys Arg 120 115 (2) INFORMATION FOR SEQ ID NO:8: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: Vg-1 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..118 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8: Arg Arg Lys Arg Ser Tyr Ser Lys Leu Pro Phe Thr Ala Ser Asn Ile

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Cys Lys Lys Arg His Leu Tyr Val Glu Phe Lys Asp Val Gly Trp Gln 25 20 Asn Trp Val Ile Ala Pro Gln Gly Tyr Met Ala Asn Tyr Cys Tyr Gly 40 35 Glu Cys Pro Tyr Pro Leu Thr Glu Ile Leu Asn Gly Ser Asn His Ala 5 60 55 50 Ile Leu Gln Thr Leu Val His Ser Ile Glu Pro Glu Asp Ile Pro Leu 75 70 65 Pro Cys Cys Val Pro Thr Lys Met Ser Pro Ile Ser Met Leu Phe Tyr 90 85 10 Asp Asn Asn Asp Asn Val Val Leu Arg His Tyr Glu Asn Met Ala Val 110 105 100 Asp Glu Cys Gly Cys Arg 115 (2) INFORMATION FOR SEQ ID NO:9: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single 20 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: Vgr-1 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..118 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9: Arg Val Ser Ser Ala Ser Asp Tyr Asn Ser Ser Glu Leu Lys Thr Ala

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Cys Arg Lys His Glu Leu Tyr Val Ser Phe Gln Asp Leu Gly Trp Gln 25 20 Asp Trp Ile Ile Ala Pro Lys Gly Tyr Ala Ala Asn Tyr Cys Asp Gly 45 40 35 Glu Cys Ser Phe Pro Leu Asn Ala His Met Asn Ala Thr Asn His Ala 5 60 55 50 Ile Val Gln Thr Leu Val His Leu Met Asn Pro Glu Tyr Val Pro Lys 75 70 65 Pro Cys Cys Ala Pro Thr Lys Leu Asn Ala Ile Ser Val Leu Tyr Phe 95 90 85 10 Asp Asp Asn Ser Asn Val Ile Leu Lys Lys Tyr Arg Asn Met Val Val 110 105 100 Arg Ala Cys Gly Cys His 115 (2) INFORMATION FOR SEQ ID NO:10: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single 20 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: OP-1 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..118 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10: Arg Met Ala Asn Val Ala Glu Asn Ser Ser Ser Asp Gln Arg Gln Ala 10 15 5 1

Cys Lys Lys His Glu Leu Tyr Val Ser Phe Arg Asp Leu Gly Trp Gln 25 Asp Trp Ile Ile Ala Pro Glu Gly Tyr Ala Ala Tyr Tyr Cys Glu Gly 40 Glu Cys Ala Phe Pro Leu Asn Ser Tyr Met Asn Ala Thr Asn His Ala 5 55 Ile Val Gln Thr Leu Val His Phe Ile Asn Pro Glu Thr Val Pro Lys 75 70 65 Pro Cys Cys Ala Pro Thr Gln Leu Asn Ala Ile Ser Val Leu Tyr Phe 90 10 85 Asp Asp Ser Ser Asn Val Ile Leu Lys Lys Tyr Arg Asn Met Val Val 105 100 Arg Ala Cys Gly Cys His 115 (2) INFORMATION FOR SEQ ID NO:11: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single 20 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: BMP-5 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..118 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11: Arg Met Ser Ser Val Gly Asp Tyr Asn Thr Ser Glu Gln Lys Gln Ala 15 10

5

Cys Lys Lys His Glu Leu Tyr Val Ser Phe Arg Asp Leu Gly Trp Gln 25 Asp Trp Ile Ile Ala Pro Glu Gly Tyr Ala Ala Phe Tyr Cys Asp Gly 40 Glu Cys Ser Phe Pro Leu Asn Ala His Met Asn Ala Thr Asn His Ala 5 60 55 Ile Val Gln Thr Leu Val His Leu Met Phe Pro Asp His Val Pro Lys 75 70 Pro Cys Cys Ala Pro Thr Lys Leu Asn Ala Ile Ser Val Leu Tyr Phe 90 85 10 Asp Asp Ser Ser Asn Val Ile Leu Lys Lys Tyr Arg Asn Met Val Val 110 105 100 Arg Ser Cys Gly Cys His 115 (2) INFORMATION FOR SEQ ID NO:12: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: 60A (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..118 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Ser Pro Asn Asn Val Pro Leu Leu Glu Pro Met Glu Ser Thr Arg Ser

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Cys Gln Met Gln Thr Leu Tyr Ile Asp Phe Lys Asp Leu Gly Trp His 20 25 Asp Trp Ile Ile Ala Pro Glu Gly Tyr Gly Ala Phe Tyr Cys Ser Gly 40 Glu Cys Asn Phe Pro Leu Asn Ala His Met Asn Ala Thr Asn His Ala 5 55 Ile Val Gln Thr Leu Val His Leu Leu Glu Pro Lys Lys Val Pro Lys 70 Pro Cys Cys Ala Pro Thr Arg Leu Gly Ala Leu Pro Val Leu Tyr His 90 10 Leu Asn Asp Glu Asn Val Asn Leu Lys Lys Tyr Arg Asn Met Ile Val 105 100 Lys Ser Cys Gly Cys His 115 15 (2) INFORMATION FOR SEQ ID NO:13: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 117 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: BMP-2 (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..117 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13: Glu Lys Arg Gln Ala Lys His Lys Gln Arg Lys Arg Leu Lys Ser Ser

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Cys Lys Arg His Pro Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn 25 Asp Trp Ile Val Ala Pro Pro Gly Tyr His Ala Phe Tyr Cys His Gly Glu Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr Asn His Ala 5 55 Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser Lys Ile Pro Lys Ala 75 70 65 Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu Tyr Leu Asp 85 10 Glu Asn Glu Lys Val Val Leu Lys Asn Tyr Gln Asp Met Val Val Glu 110 105 100 Gly Cys Gly Cys Arg 115 (2) INFORMATION FOR SEQ ID NO:14: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 117 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: BMP-4 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..117 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14: Arg Ser Pro Lys His His Ser Gln Arg Ala Arg Lys Lys Asn Lys Asn

5

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Cys Arg Arg His Ser Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn 20 Asp Trp Ile Val Ala Pro Pro Gly Tyr Gln Ala Phe Tyr Cys His Gly Asp Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr Asn His Ala 5 55 Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser Ser Ile Pro Lys Ala 75 70 65 Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu Tyr Leu Asp 90 10 85 Glu Tyr Asp Lys Val Val Leu Lys Asn Tyr Gln Glu Met Val Val Glu 110 105 100 Gly Cys Gly Cys Arg 115 (2) INFORMATION FOR SEQ ID NO:15: 15 (1) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: DPP (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..118 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15: Lys Arg His Ala Arg Arg Pro Thr Arg Arg Lys Asn His Asp Asp Thr

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Cys Arg Arg His Ser Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asp 25 20 Asp Trp Ile Val Ala Pro Leu Gly Tyr Asp Ala Tyr Tyr Cys His Gly 40 35 Lys Cys Pro Phe Pro Leu Ala Asp His Phe Asn Ser Thr Asn His Ala 5 55 Val Val Gln Thr Leu Val Asn Asn Met Asn Pro Gly Lys Val Pro Lys 75 70 65 Ala Cys Cys Val Pro Thr Gln Leu Asp Ser Val Ala Met Leu Tyr Leu 90 85 10 Asn Asp Gln Ser Thr Val Val Leu Lys Asn Tyr Gln Glu Met Thr Val 105 100 Val Gly Cys Gly Cys Arg 115 (2) INFORMATION FOR SEQ ID NO:16: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 119 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: BMP-3 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..119 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Gln Thr Leu Lys Lys Ala Arg Arg Lys Gln Trp Ile Glu Pro Arg Asn

Cys Ala Arg Arg Tyr Leu Lys Val Asp Phe Ala Asp Ile Gly Trp Ser 25 Glu Trp Ile Ile Ser Pro Lys Ser Phe Asp Ala Tyr Tyr Cys Ser Gly 40 35 Ala Cys Gln Phe Pro Met Pro Lys Ser Leu Lys Pro Ser Asn His Ala 5 55 Thr Ile Gln Ser Ile Val Arg Ala Val Gly Val Val Pro Gly Ile Pro 75 70 65 Glu Pro Cys Cys Val Pro Glu Lys Met Ser Ser Leu Ser Ile Leu Phe 85 10 Phe Asp Glu Asn Lys Asn Val Val Leu Lys Val Tyr Pro Asn Met Thr 105 110 100 Val Glu Ser Cys Ala Cys Arg 115 (2) INFORMATION FOR SEQ ID NO:17: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 115 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: MIS (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..115 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17: Pro Gly Arg Ala Gln Arg Ser Ala Gly Ala Thr Ala Ala Asp Gly Pro 15 10 1

Cys Ala Leu Arg Glu Leu Ser Val Asp Leu Arg Ala Glu Arg Ser Val 25 20 Leu Ile Pro Glu Thr Tyr Gln Ala Asn Asn Cys Gln Gly Val Cys Gly 40 35 Trp Pro Gln Ser Asp Arg Asn Pro Arg Tyr Gly Asn His Val Val Leu 5 55 50 Leu Leu Lys Met Gln Ala Arg Gly Ala Ala Leu Ala Arg Pro Pro Cys 65 70 Cys Val Pro Thr Ala Tyr Ala Gly Lys Leu Leu Ile Ser Leu Ser Glu 95 90 10 85 Glu Arg Ile Ser Ala His His Val Pro Asn Met Val Ala Thr Glu Cys 110 105 100 Gly Cys Arg 115 15 (2) INFORMATION FOR SEQ ID NO:18: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 121 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: Inhibin alpha (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..121 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18: Leu Arg Leu Leu Gln Arg Pro Pro Glu Glu Pro Ala Ala His Ala Asn 15 5 10

Cys His Arg Val Ala Leu Asn Ile Ser Phe Gln Glu Leu Gly Trp Glu 25 20 Arg Trp Ile Val Tyr Pro Pro Ser Phe Ile Phe His Tyr Cys His Gly 40 Gly Cys Gly Leu His Ile Pro Pro Asn Leu Ser Leu Pro Val Pro Gly 5 55 Ala Pro Pro Thr Pro Ala Gln Pro Tyr Ser Leu Leu Pro Gly Ala Gln 75 70 65 Pro Cys Cys Ala Ala Leu Pro Gly Thr Met Arg Pro Leu His Val Arg 90 10 85 Thr Thr Ser Asp Gly Gly Tyr Ser Phe Lys Tyr Glu Thr Val Pro Asn 110 105 100 Leu Leu Thr Gln His Cys Ala Cys Ile 115 120 (2) INFORMATION FOR SEQ ID NO:19: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 121 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: Inhibin betaA (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..121 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Arg Arg Arg Arg Gly Leu Glu Cys Asp Gly Lys Val Asn Ile Cys

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Cys Lys Lys Gln Phe Phe Val Ser Phe Lys Asp Ile Gly Trp Asn Asp 25 20 Trp Ile Ile Ala Pro Ser Gly Tyr His Ala Asn Tyr Cys Glu Gly Glu 40 35 Cys Pro Ser His Ile Ala Gly Thr Ser Gly Ser Ser Leu Ser Phe His 5 50 55 Ser Thr Val Ile Asn His Tyr Arg Met Arg Gly His Ser Pro Phe Ala 75 70 65 Asn Leu Lys Ser Cys Cys Val Pro Thr Lys Leu Arg Pro Met Ser Met 95 90 85 10 -Leu Tyr Tyr Asp Asp Gly Gln Asn Ile Ile Lys Lys Asp Ile Gln Asn 110 105 100 Met Ile Val Glu Glu Cys Gly Cys Ser 120 115 (2) INFORMATION FOR SEQ ID NO:20: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single 20 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: Inhibin betaB (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..120 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Arg Ile Arg Lys Arg Gly Leu Glu Cys Asp Gly Arg Thr Asn Leu Cys

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Cys Arg Gln Gln Phe Phe Ile Asp Phe Arg Leu Ile Gly Trp Asn Asp 25 Trp Ile Ile Ala Pro Thr Gly Tyr Tyr Gly Asn Tyr Cys Glu Gly Ser 40 Cys Pro Ala Tyr Leu Ala Gly Val Pro Gly Ser Ala Ser Ser Phe His 5 55 Thr Ala Val Val Asn Gln Tyr Arg Met Arg Gly Leu Asn Pro Gly Thr 70 Val Asn Ser Cys Cys Ile Pro Thr Lys Leu Ser Thr Met Ser Met Leu 90 10 Tyr Phe Asp Asp Glu Tyr Asn Ile Val Lys Arg Asp Val Pro Asn Met 100 105 Ile Val Glu Glu Cys Gly Cys Ala 115 (2) INFORMATION FOR SEQ ID NO:21: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 114 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single 20 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: TGF-betal (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..114 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21: Arg Arg Ala Leu Asp Thr Asn Tyr Cys Phe Ser Ser Thr Glu Lys Asn 15 5 10

Cys Cys Val Arg Gln Leu Tyr Ile Asp Phe Arg Lys Asp Leu Gly Trp 25 Lys Trp Ile His Glu Pro Lys Gly Tyr His Ala Asn Phe Cys Leu Gly 40 Pro Cys Pro Tyr Ile Trp Ser Leu Asp Thr Gln Tyr Ser Lys Val Leu 5 55 Ala Leu Tyr Asn Gln His Asn Pro Gly Ala Ser Ala Ala Pro Cys Cys 70 65 Val Pro Gln Ala Leu Glu Pro Leu Pro Ile Val Tyr Tyr Val Gly Arg 10 Lys Pro Lys Val Glu Gln Leu Ser Asn Met Ile Val Arg Ser Cys Lys 110 105 100

15 (2) INFORMATION FOR SEQ ID NO:22:

Cys Ser

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 114 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
- 20 (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (vii) IMMEDIATE SOURCE:
    - (B) CLONE: TGF-beta2
  - (ix) FEATURE:

25 (A) NAME/KEY: Protein

(B) LOCATION: 1..114

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Lys Arg Ala Leu Asp Ala Ala Tyr Cys Phe Arg Asn Val Gln Asp Asn 1 5 10 15

Cys Cys Leu Arg Pro Leu Tyr Ile Asp Phe Lys Arg Asp Leu Gly Trp 25 Lys Trp Ile His Glu Pro Lys Gly Tyr Asn Ala Asn Phe Cys Ala Gly 40 Ala Cys Pro Tyr Leu Trp Ser Ser Asp Thr Gln His Ser Arg Val Leu 5 55 Ser Leu Tyr Asn Thr Ile Asn Pro Glu Ala Ser Ala Ser Pro Cys Cys 75 70 65 Val Ser Gln Asp Leu Glu Pro Leu Thr Ile Leu Tyr Tyr Ile Gly Lys 90 85 10 Thr Pro Lys Ile Glu Gln Leu Ser Asn Met Ile Val Lys Ser Cys Lys 105 100 Cys Ser (2) INFORMATION FOR SEQ ID NO:23: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 114 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: TGF-beta3 (ix) FEATURE: (A) NAME/KEY: Protein 25 (B) LOCATION: 1..114 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23: Lys Arg Ala Leu Asp Thr Asn Tyr Cys Phe Arg Asn Leu Glu Glu Asn

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Cys Cys Val Arg Pro Leu Tyr Ile Asp Phe Arg Gln Asp Leu Gly Trp 25 20 Lys Trp Val His Glu Pro Lys Gly Tyr Tyr Ala Asn Phe Cys Ser Gly 40 35 Pro Cys Pro Tyr Leu Arg Ser Ala Asp Thr Thr His Ser Thr Val Leu 5 50 55 Gly Leu Tyr Asn Thr Leu Asn Pro Glu Ala Ser Ala Ser Pro Cys Cys 75 70 65 Val Pro Gln Asp Leu Glu Pro Leu Thr Ile Leu Tyr Tyr Val Gly Arg 90 85 10 Thr Pro Lys Val Glu Gln Leu Ser Asn Met Val Val Lys Ser Cys Lys 105 100 Cys Ser

- 15 (2) INFORMATION FOR SEQ ID NO:24:
  - (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 116 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

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- (ii) MOLECULE TYPE: protein
- (vii) IMMEDIATE SOURCE:

(B) CLONE: TGF-beta4

(ix) FEATURE:

(A) NAME/KEY: Protein

(B) LOCATION: 1..116

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Arg Arg Asp Leu Asp Thr Asp Tyr Cys Phe Gly Pro Gly Thr Asp Glu
1 5 10 15

Lys Asn Cys Cys Val Arg Pro Leu Tyr Ile Asp Phe Arg Lys Asp Leu 25 20 Gln Trp Lys Trp Ile His Glu Pro Lys Gly Tyr Met Ala Asn Phe Cys 40 Met Gly Pro Cys Pro Tyr Ile Trp Ser Ala Asp Thr Gln Tyr Thr Lys 5 60 50 55 Val Leu Ala Leu Tyr Asn Gln His Asn Pro Gly Ala Ser Ala Ala Pro 75 70 65 Cys Cys Val Pro Gln Thr Leu Asp Pro Leu Pro Ile Ile Tyr Tyr Val 90 10 85 Gly Arg Asn Val Arg Val Glu Gln Leu Ser Asn Met Val Val Arg Ala 110 105 Cys Lys Cys Ser 115 (2) INFORMATION FOR SEQ ID NO:25: 15 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 114 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 20 (ii) MOLECULE TYPE: protein (vii) IMMEDIATE SOURCE: (B) CLONE: TGF-beta5 (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..114 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

Lys Arg Gly Val Gly Gln Glu Tyr Cys Phe Gly Asn Asn Gly Pro Asn

5

10

Cys Cys Val Lys Pro Leu Tyr Ile Asn Phe Arg Lys Asp Leu Gly Trp 20 25 30

Lys Trp Ile His Glu Pro Lys Gly Tyr Glu Ala Asn Tyr Cys Leu Gly 45

Asn Cys Pro Tyr Ile Trp Ser Met Asp Thr Gln Tyr Ser Lys Val Leu 50

Ser Leu Tyr Asn Gln Asn Asn Pro Gly Ala Ser Ile Ser Pro Cys Cys 65 70 75 80

Val Pro Asp Val Leu Glu Pro Leu Pro Ile Ile Tyr Tyr Val Gly Arg 85 90 95

Thr Ala Lys Val Glu Gln Leu Ser Asn Met Val Val Arg Ser Cys Asn 100 105 110

Cys Ser

5

10

20

25

## 15 (2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 454 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vii) IMMEDIATE SOURCE:

(B) CLONE: HUMAN GDF-9

(ix) FEATURE:

(A) NAME/KEY: Protein

(B) LOCATION: 1..454

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

	Met 1	Ala	Arg	Pro	Asn 5	Lys	rne	Leu	Leu	10	THE	Oy 3	0,5		15	**P
	Leu	Cys	Phe	Pro 20	Ile	Ser	Leu	Gly	Ser 25	Gln	Ala	Ser	Gly	Gly 30	Glu	Ala
5	Gln	lle	Ala 35	Ala	Ser	Ala		Leu 40	Glu	Ser	Gly	Ala	Met 45	Pro	Trp	Ser
	Leu	Leu 50	Gln	His	Ile	Asp	Glu 55	Arg	Asp	Arg	Ala	Gly 60	Leu	Leu	Pro	Ala
10	Leu 65	Phe	Lys	Val	Leu	Ser 70	Val	Gly	Arg	Gly	Gly 75	Ser	Pro	Arg	Leu	Gln 80
	Pro	Asp	Ser	Arg	Ala 85	Leu	His	Tyr	Met	Lys 90	Lys	Leu	Tyr	Lys	Thr 95	Tyr
	Ala	Thr	Lys	Glu 100	Gly	Ile	Pro	Lys	Ser 105	Asn	Arg	Ser	His	Leu 110	Tyr	Asn
15	Thr	Val	Arg 115		Phe	Thr	Pro	Cys 120	Thr	Arg	His	Lys	Gln 125	Ala	Pro	Gly
	Asp	Gln 130	Val	Thr	G1y	Ile	Leu 135	Pro	Ser	Val	Glu	Leu 140		Phe	Asn	Leu
20	Asp 145		Ile	Thr	Thr	Val 150	Glu	His	Leu	Leu	Lys 155		Val	Leu	Leu	Tyr 160
	Asn	Ile	Asn	Asn	Ser 165		Ser	Phe	Ser	Ser 170		. Val	Lys	Cys	Val 175	
	Asn	Leu	Met	11e		Glu	Pro	Lys	Ser 185		Ser	Arg	Thr	190	Gly	Arg
25	Ala	Pro	Tyr 195		Phe	Thr	Phe	200		c Gln	Phe	: Glu	205		Lys	Lys
	His	210		ıle	Glr	ılle	Asp 215		Thi	r Ser	Leu	220		n Pro	Lev	ı Val
30	Ala 225		Ast	n Lys	Arg	3 Ser 230		e His	s Me	t Ser	235		n Phe	e Thi	Cy:	Met 240

	Lys	Asp	Gln	Leu	Glu : 245	His	Pro	Ser	Ala	Gln 250	Asn	Gly	Leu	Phe	Asn 255	Met
	Thr	Leu	Val	Ser 260	Pro	Ser	Leu	Ile	Leu 265	Tyr	Leu	Asn	Asp	Thr 270	Ser	Ala
5	Gln	Ala	Tyr 275	His	Ser	Trp	Tyr	Ser 280	Leu	His	Tyr	Lys	Arg 285	Arg	Pro	Ser
	Gln	Gly 290	Pro	Asp	Gln	Glu	Arg 295	Ser	Leu	Ser	Ala	Tyr 300	Pro	Val	Gly	Glu
10	Glu 305	Ala	Ala	Glu	Asp	Gly 310	Arg	Ser	Ser	His	His 315	Arg	His	Arg	Arg	Gly 320
	Gln	Glu	Thr	Val	Ser 325	Ser	Glu	Leu	Lys	Lys 330	Pro	Leu	Gly	Pro	Ala 335	Ser
	Phe	Asn	Leu	Ser 340		Tyr	Phe	Arg	Gln 345		Leu	Leu	Pro	Gln 350	Asn	Glu
15	Cys	Glu	Leu 355		Asp	Phe	Arg	Leu 360		Phe	Ser	Gln	Leu 365	Lys	Trp	Asp
	Asn	370		. Val	Ala	Pro	His		Tyr	: Asn	Pro	Arg 380	Tyr	Cýs	Lys	Gly
20	Asp 385		Pro	Arg	; Ala	Val 390		His	Arg	g Tyr	Gly 395	Ser	Pro	Val	His	400
	Met	: Val	Gl:	n Asr	11e 405		туі	c Glu	ı Ly:	s Leu 410	ı Ası	Ser	Ser	· Val	415	Arg
	Pro	o Sei	r Cy:	s Val		Ala	a Ly:	s Ty	r Se:		Let	ı Sei	r Val	43(	ı Thi	r Ile
25	Gl	u Pr	o As 43		y Sei	r Il	e Al	a Ty:		s Gl	u Ty:	r Gl	u Ası 44!	p Met	t Il	e Ala
	Th	r Ly 45		s Th	r Cy	s Ar	g									

## **CLAIMS**

- 1. Substantially pure growth differentiation factor-9 (GDF-9) and functional fragments thereof.
- 2. An isolated polynucleotide sequence encoding the GDF-9 polypeptide of claim 1.
- 3. The polynucleotide sequence of claim 2, wherein the polynucleotide is isolated from a mammalian cell.
- 4. The polynucleotide of claim 3, wherein the mammalian cell is selected from the group consisting of mouse, rat, and human cell.
- 5. An expression vector including the polynucleotide of claim 2.
- 6. The vector of claim 5, wherein the vector is a plasmid.
- 7. The vector of claim 5, wherein the vector is a virus.
- 8. A host cell stably transformed with the vector of claim 5.
- 9. The host cell of claim 8, wherein the cell is prokaryotic.
- 10. The host cell of claim 8, wherein the cell is eukaryotic.
- 11. Antibodies reactive with the polypeptide of claim 1 or fragments thereof.
- 12. The antibodies of claim 11, wherein the antibodies are polyclonal.

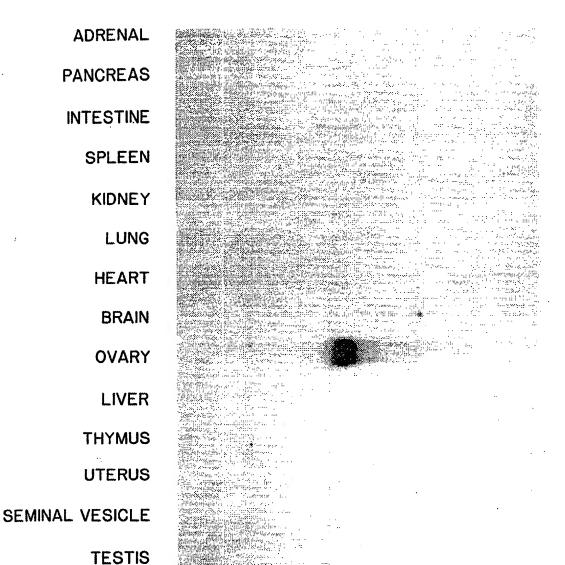
- 13. The antibodies of claim 11, wherein the antibodies are monoclonal.
- 14. A method of detecting a cell proliferative disorder comprising contacting the antibody of claim 11 with a specimen of a subject suspected of having a GDF-9 associated disorder and detecting binding of the antibody.
- 15. The method of claim 14, wherein the cell proliferative disorder is an ovarian tumor.
- 16. The method of claim 14, wherein the detecting is in vivo.
- 17. The method of claim 16, wherein the antibody is detectably labeled.
- 18. The method of claim 17, wherein the detectable label is selected from the group consisting of a radioisotope, a fluorescent compound, a bioluminescent compound and a chemiluminescent compound.
- 19. The method of claim 14, wherein the detection is in vitro.
- 20. The method of claim 19, wherein the antibody is detectably labeled.
- 21. The method of claim 20, wherein the label is selected from the group consisting of a radioisotope, a fluorescent compound, a bioluminescent compound, a chemoluminescent compound and an enzyme.
- 22. A method of treating a cell proliferative disorder associated with expression of GDF-9, comprising contacting the cells with a reagent which suppresses the GDF-9 activity.

- 23. The method of claim 22, wherein the reagent is an anti-GDF-9 antibody.
- 24. The method of claim 22, wherein the reagent is a GDF-9 antisense sequence.
- 25. The method of claim 22, wherein the cell proliferative disorder is an ovarian tumor.
- 26. The method of claim 22, wherein the reagent which suppresses GDF-9 activity is introduced to a cell using a vector.
- 27. The method of claim 26, wherein the vector is a colloidal dispersion system.
- 28. The method of claim 27, wherein the colloidal dispersion system is a liposome.
- 29. The method of claim 28, wherein the liposome is essentially target specific.
- 30. The method of claim 29, wherein the liposome is anatomically targeted.
- 31. The method of claim 29, wherein the liposome is mechanistically targeted.
- 32. The method of claim 31, wherein the mechanistic targeting is passive.
- 33. The method of claim 31, wherein the mechanistic targeting is active.

- 34. The method of claim 33, wherein the liposome is actively targeted by coupling with a moiety selected from the group consisting of a sugar, a glycolipid, and a protein.
- 35. The method of claim 34, wherein the protein moiety is an antibody.
- 36. The method of claim 35, wherein the vector is a virus.
- 37. The method of claim 36, wherein the virus is an RNA virus.
- 38. The method of claim 37, wherein the RNA virus is a retrovirus.
- 39. The method of claim 38, wherein the retrovirus is essentially target specific.

10.5 d PLACENTA

- 1.7 kb



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# FIG.2a

rc 660	720	CC 780	CT 840	ST 900	3/	15 096 IX	<u>_</u>	4A 1020		CA 1080	~	3G 1140	(1)
	N F T	CTC	TCTC		о О	SCCZ	_	TCZ	*	\GTC	S	AAC	×
TGA	TAT		AGT	GTC	<b>8</b>	AAG	<b>A</b>	ACT	) <u>I</u> I,	TTP		CTZ	<b>X</b> ,
ATG	OL A		900	CC K	A	AGA	A X O	AAT	E Y F	GTT	14	ACT	O
ATGGATTGAGATTGATGTGAC W I E I D V T	AGCATTCACCTGTCTGT	GGAGTGTTTAGCATGCCTCTCTCAGTGCC	CAGGCCTACCACTCTTGGCAC	crs	A	GGC	O <sup>i</sup>	SCG	<u>ш</u>	AGTGTGAACTCCATGACTTCAGACTGAGTTTTAGTCA	CELHDFRLSF	GGT	P R Y C
AGA	TGT	CTC 1.	ACT	16G	A	GAG	ਹ _	TCA	N L S	GAC	J	CTA	æ
ITG.	ACC'	IGC.	ACC.	GTG	>	3TC	R	ACC	L	ICA	K	ACC	
3GA' I	rTC, H	GCA'	CCT.	, SCG	H P G Q A G V A	3CC	R R R G	ICA	Z	ACT	[L	ACA	R Y N
3ATK W	3CA'	TTA ?	AGG(	AGG(	Ø	3GC	X	CT	দে	ATG.	Ω	3GT	×
ACAG. R	3GA(	IGT' F	$\frac{1}{2}$	3CCX	Ø	$\sum_{i=1}^{n}$		ATC	ഗ	rcc.	H	<b>ACA</b>	ĸ
AACAC H	AGAGG R	SAG	AAGCACC	() ()	U	CTC	Д	SAG	·L T A	<b>JAC</b>	J	CCZ	Н
AAGAAA K	CGAG E	) (CGC	ZAAC 7	TCC	Δ,	ATC	Ŋ	TAC	H	TG7	ы	$\mathcal{CC}$	ፈ
GAZ K	CAGG	AGGAC	TCAC T	CC	H	AAC	ĸ	TC1	Ĥ	\GTC		CGC	Ø
CCC	CTC	AGZ E	CGC	ACZ	Ø	CG2	Ш	ACT	IJ	CGA	ы	CGI	>
ATTCAC F T	CGAC	ညည်း			J	\GG1	E V E	BGCC	Д	AAA	E Z O	GAT	H
ATT	CTCCTTCAGCCCCTAGTGAC	TCCACAAAAGACCAGGTGCCAGAG	CTCATCTTGTATCTCAACGACACA	200	TWRPLQ	TG		000	EAKGPL	CCA		CTC	LKWDNWIV
AGCACCGTACTC! A P Y S	CCT	200	STA *	GAG	ĸ	TAC	E E A T	GAA	×	CCC	F L F P	CAA	Z
GTA	) )	AGA	CTT	CIG	3	AGC	Ø	AGC	K	TTT	ഥ	GGA	Ω
ACC	TCA	AAA X	CAT	CAC	۲	GGA	Ш	CGA	ы	TCT	J	ATG	3
AGC	CCT I.	CAC	CCT T.	$\tilde{\mathrm{GTC}}$	ഗ	AGA	ы	CTC	ഗ	GTT	Ŀ	CAA	×
AAGAGCACCGTACTCACCCTGAAGAACACAGATGGATTGAGATTGATGTGACCTC R A P Y S F T L K K H R W I E I D V T S	CCTCCTTCAGCCCCCTAGTGACCTCCAGCGAGGAGCATTCACCTGTCTGT	ATGCACAAAAGACCAGGAGGACGGAGTGTTTAGCATGCCTCTCTCAGTGCCTCC	Trecereateraterateraceaeaeaecaeceaegectaceaererregeagrerer	TCAGTCCACCTGGAGGCCTTTACAGCATCCCGGCCAGGCCGGTGTGGCTGCCCGTCCCGT	Q	GAAAGAGGAAGCTACTGAGGTGGAAAGATCTCCCCGGCGCCGTCGAGGGCAGAAAGCCAT	×	CCGCTCCGAAGCGAAGGGGCCACTTCTTACAGCATCCTTCAACCTCAGCGAATACTTCAA	æ	ACAGITICITITICCCCCAAAACG	Ø	GCTCAAATGGGACAACTGGATCGTGGCCCCCGCACAGGTACAACCCTAGGTACTGAAAGG	IJ
601	661	721	781	841		901		961		1021		1081	
				S	UB	STI	TU	TE	SI	HE	ΞT		

## **FIG.2**t

### **FIG.20**

HGEOPFSMTTYLNS KGDOPRAVRHRYGS	CGC CAL PVALSGSGGP YGE CPY PLTEILNG DGE SFPLNAHMNA CGC CAF PLNAHMNA CGC CAF PLADHLNS HGE CPF PLADHLNS HGC CPF PLADHLNS HGC CENPELADHLNS HGC CENPELADHLNS HGC CENPELADHLNS HGC CENPERSIRP CGC CENPERSIRP CGC CENT I PPNLSLPV EGE CPSH I AGTSGSSL EGS CPSH I AGTSGSSL	MGP@PYIWSAD
EXERCISE SECTION SECTI	╸meseseseseseseseseseseseseseseseseseses	
RNF CHRHQLFINF-QDLGWHKWVIAPKGFMANYCHGEOPFSMTTYLNS- QNECELHDFRLSF-SQLKWDNWIVAPHRYNPRYCKGDOPRAVRHRYGS-	PRRDAEPVLGGGPGGACARRILYVSF-REVGWHRWVIAPRGFLANY  RKKRSYSKLPFTASNICKRHLYVSF-REVGWHRWVIAPRGFLANY  RVSSASDYNSSELKTACRKHLLYVSF-QDLGWQDWIIAPRGYAANY  RVSSASDYNSSELKTACRKHELYVSF-QDLGWQDWIIAPRGYAANY  RMANVAENSSDQRQACKKHELYVSF-RDLGWQDWIIAPRGYAAYY  CEGE AFPLNAHMNASPNNYBESQRQACKKHELYVSF-RDLGWQDWIIAPRGYAAYY  CEGE AFPLNAHMNASPNNYBESQTRSCQMCHLIYVSF-RDLGWQDWIIAPRGYAAYY  SPNNYPLLEPMESTRSCQMCHLIYVSF-RDLGWWDWIIAAPRGYGAFY  CERRQAKHKQRKRLKSSCKRHPLYVDF-SDVGWNDWIIVAPPGYGAFY  CRACKRORKNKNCRRHSLYVDF-SDVGWNDWIIVAPPGYQAFY  CRACKRORKDDTCRRHSLYVDF-SDVGWNDWIIVAPPGYQAFY  CRACKRORKDDTCRRHSLYVDF-SDVGWNDWIIVAPPGYQAFY  CRACKRORKNNCRRYLKVDF-ADIGWSEWIISPKSFIDAYY  CRACKRORYNICKRQFFVSF-KDIGWNDWIIAPSGYHANY  CRACKRORYNICKRQFFVSF-KDIGWNDWIIAPSGYHANY  CRACKROLECDGRYNICKRQFFVSF-KDIGWNDWIIAPSGYHANY  CRALLDTNYCFSSTEKNCVRQLYIDFRKDLGWK-WIHEPRGYNANFCGGEPPYLMSSD  KRALLDTNYCFSSTEKNCVRQLYIDFRKDLGWK-WIHEPRGYNANFGGEPPYLMSSD  KRALLDTNYCFSSTEKNCVRPLYNDF-WHEPRGYYANNFGGEPPYLMSSD  KRALLDTNYCFRNIFFN CVRPLYIDFREDLGWK-WHEPRGYYANNFGGEPPYLMSSD	RRDLDTDYCFGPGTDEKNCCVRPLYIDFRKDLOWK-WIHEPKGYMANFOMGPPYIWSAD- KRGVGQEYCFGNNGPNCCVKPLYINFRKDLGWK-WIHEPKGYEANYCLGNGPYIWSMD-
¥ 2	- GGA - SNH - KTA - KOA - TROA - TROA - DDA - DDA - DGB - DGB - NH - DGB - NH - DGB - DGB - DGB - DGB - DGB - DGB NH - NH - NH - NH - NH - NH - NH -	NA
KRRAAISVPKGFCR	PRRDAEPVLGGGPG RRKRSYSKLPFTAS RVSSASDYNSSELK RWANVAENSSSDQR SPNNVPLLEPMEST EKRQAKHKQRKRLK RSPKHHSQRARKKN KRHARRPTRRKNHD QTLKKARRKQWIEP PGRAQRSAGATAAD LRLLQRPPEEPAAH RRRRGLECDGKV RIRKRGLECDGKV RIRKRGLECDGKY	RRDLDTDYCFGPGTDE KRGVGQEYCFGNNG
	α ßa ßb	
GDF-3 GDF-9	GDF-1 Vg-1 Vgr-1 OP-1 BMP-2 BMP-4 DPP BMP-4 DPP Inhibin Inhibin TGF-61 TGF-62	TGF-84 TGF-85

FIG.38

() C	C C I I	ESESE	4 K K	erere KKH	SAS	
SNYAFMQALMHMADPKVPKAVCVPTKLSPISMLYQ-DSDKNVILRHYEDMVVDEC	PALNHAVLRALMHA AAPGAADL PCOV - PARLSPISVLFF - DNSDNVVLRQYEDMVVDE CO - SNHAILQTLVHS - IEPEDIPLPCOV - PTKMSPISMLFY - DNNDNVVLRHYENMAVDE CO - TNHAIVQTLVHL - MNPEYVPKPCOA - PTKLNAISVLYF - DDNSNVILKKYRNMVVRACOA - PTKLNAISVLYF - PTKLNAI	INHAIVQTLVHLMFPDHVPKPCAPTKLNAISVLYF-DDSSNVILKKYRNMVVRSC TNHAIVQTLVHLLEPKKVPKPCAPTRLGALPVLYH-LNDENVNLKKYRNMIVKSC TNHAIVQTLVHLLEPKKVPKPCAPTRLGALPVLYH-LNDENVNLKKYRNMIVKSC	TNHAIVQTLVNSVNSKIPKACVPTELSAISMLIL-DENEKVVLKNYQEMVVEGC TNHAIVQTLVNSVNSSIPKACVPTELSAISMLYL-DEYDKVVLKNYQEMVVEGC TNHAVVQTLVNNMNPGKVPKACVPTQLDSVAMLYL-NDQSTVVLKNYQEMTVVGC	SNHATIQSIVRA-VGVVPGIPEPCOVPEKMSSLSILFF-DENKNVVLKVYPNMTVESOV GNHVVLLLKMQARGAALARPPCOVPTAYAGKLLISLSEERISAHHVPNMVATEC pgapptpaopysllpgaopcaalpgtmRplhvRTTSDGGYSFKYETVPNLLTOHOV	SFHSTVINHYRMRGHSPFANLKSCVPTKLRPMSMLYY-DDGQNIIKKDIQNMIVEEC	TQYSKVLALYNQHNPGASAAPGGVPQALEPLPIVYY-VGRKPKV-EQLSNMIVKSGKGSTQHSRVLSLYNTINPEASASPGGVSQDLEPLTILYY-IGKTPKI-EQLSNMIVKSGKGSTTHSTVLGLYNTLNPEASASPGGVPQDLEPLTILYY-VGRTPKV-EQLSNMVVKSGKGSTQYTKVLALYNQHNPGASAAPGGVPQTLDPLPIIYY-VGRNVRV-EQLSNMVVRAGKGSTQYTKVLALYNQNNPGASISPGGVPDVLEPLPIIYY-VGRNVRV-EQLSNMVVRSGNGS
GDF-3 GDF-9	GDF-1 Vg-1 Vgr-1		BMP-2 BMP-4 DPP	BMP-3 MIS Tabibin Q	i d d	TGF-81 TGF-82 TGF-84 TGF-85

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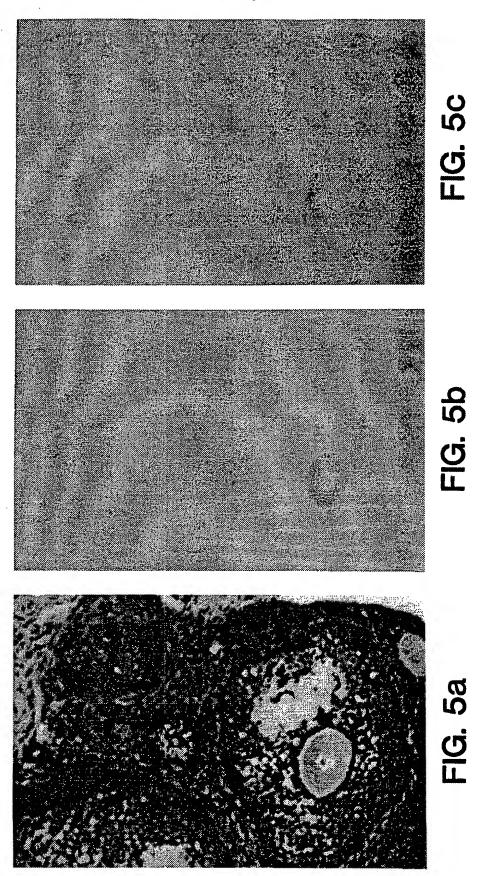
### FIG.3b

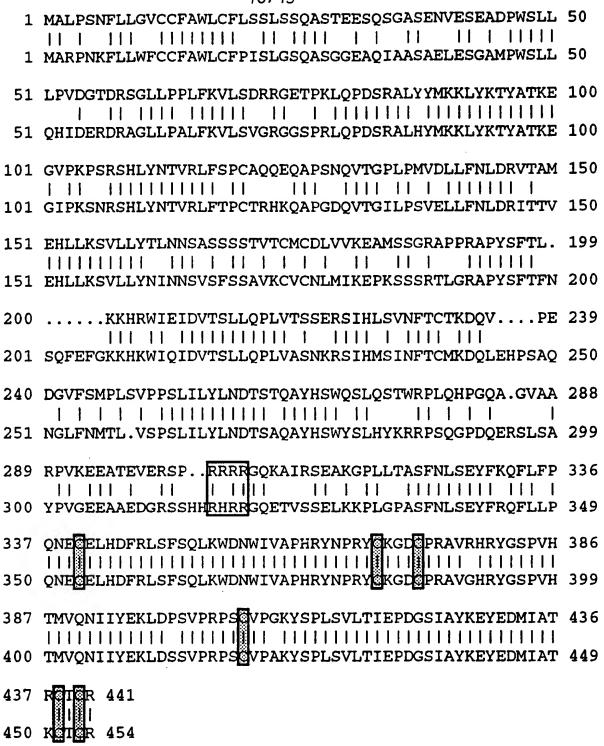
BMP-2	<b>~</b>	•					61	57	00		
BMP-2	53	33	42	28	61	60					
ВМР-4	20	34	43	26	9	28	59 57	54 54	92 74	100 75	- 100
BMP-3	47 42	32 29	41 42	48 49	59 44	58 42	57 43	54 41	74 48		00 43
MIS	77	12	34	30	24	27	24 2	25 2	27 2	27 2	25 1
INHIBIN βA INHIBIN α	25 42	27 30	23 37	22 44	25 44	24 43	24 43	24 36	22 42	22 41	19 39
INHIBIN BB	41 3	31 23	35 33	37 34	41 35	42 34	37 34	39 38	42 35	42 34	42 35
TGF-β2	36 31 3	25	32	36	37	38	35	39	34	4 33 3	5 35 3
TGF-β4	32 33	25 22	33 34	37 32	39 39	38 37	36 36	40 38	36 33	5 32	5 33
TGF- <b>β</b> 5	36	25	32	34	37	36	36	36	35	33	35

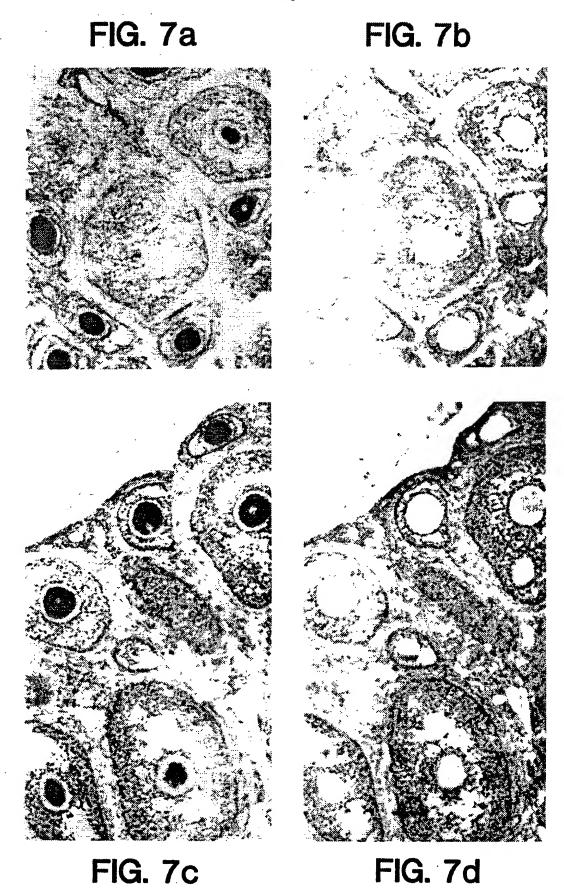
FIG.4a

				•							_
TGF- <b>β</b> 5	30	<b>5</b> 6	24	36	28	82	10	73	79	100	
TGF−β4	27	53	24	33	30	86	68	74	100	1	
TGF- <b>β</b> 3	32	25	24	36	37	78	82	100	1	1	
TGF- <b>β</b> 2	32	23	22	37	34	74	100	1	1	•	
TGF− <i>β</i> Ι	32	28	23	41	32	100	ı	1	. 1	•	!
INHIBIN &B	37	25	25	63	100	1	ı	ı	ı	1	j
INHIBIN PA	36	24	56	100	1	,	1	t	ŧ	ı	
INHIBIN a	29	18	100	1	1	١	1	ı	1	•	
MIS	30	100	i	ı	1	•	ı	ı	ı	ı	
BMP-3	100	ı	i	•	ı	1	1		٠	•	
DPP	ı	ı	ı	1	ı	1	1	ı	i	i	
BMP-4	•	1	ı	•	ı	1	1	ı	ı	ı	<b>4</b> b
BMP-2	ı	1	ı	ı	1	1	ı	1	ı	1	<b>G.4</b>
60A	ı	1	1	ı	ı	1	ı	ı	ı	ı	Ц
BMP-5	1	1	•	t	1	ı	ı	ı	ı	ı	
OP-I	1	i	•	,	1	ı	•	ı	1	ı	
Vgr-I	ı	1	ı	•	1	ı	1	•	ı	ı	
Vg-1	1	i	ı	•	1	ı	ı	ı	ı	ı	
GDF-I	1	ı	i	1	t	ı	ı	1	ı	ı	
GDF-9	1	1	ι	•	1	•	ı	1	ı	ı	
GDF-3	1	ı	t	•	•	1	1	•	1	ı	
	BMP-3	MIS	INHIBIN d	INHIBIN BA	INHIBIN BB	TGF-B1	TGF-82	TGF-83	TGF-84	TGF-\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	

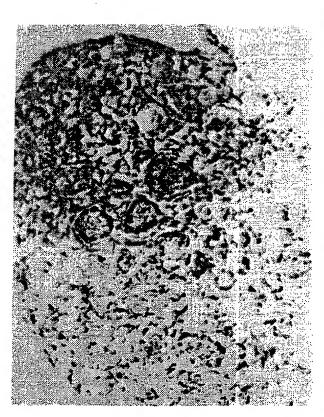
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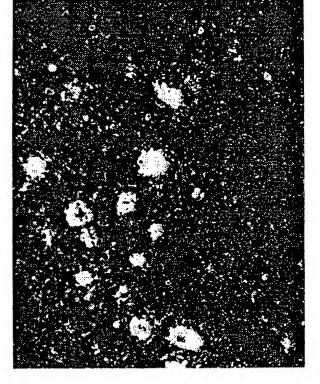
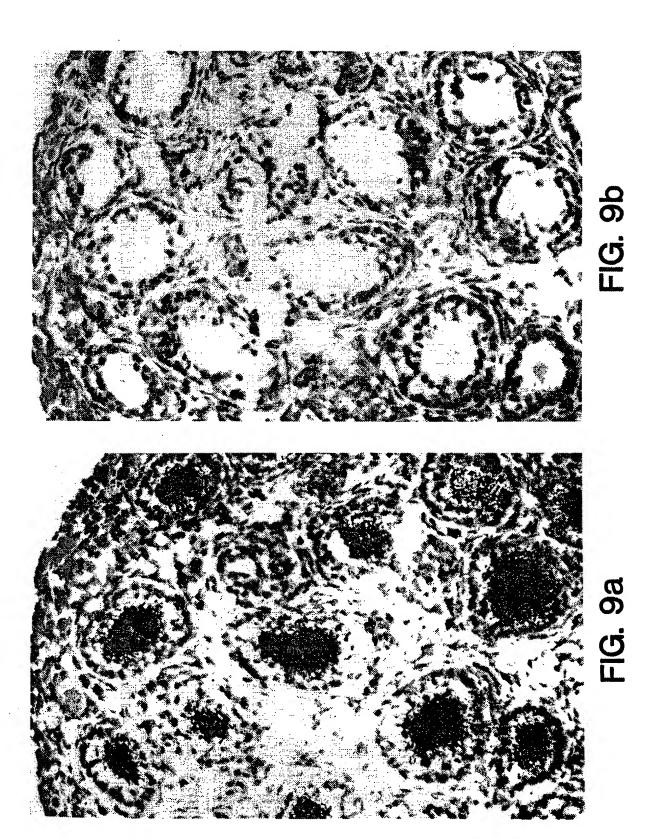
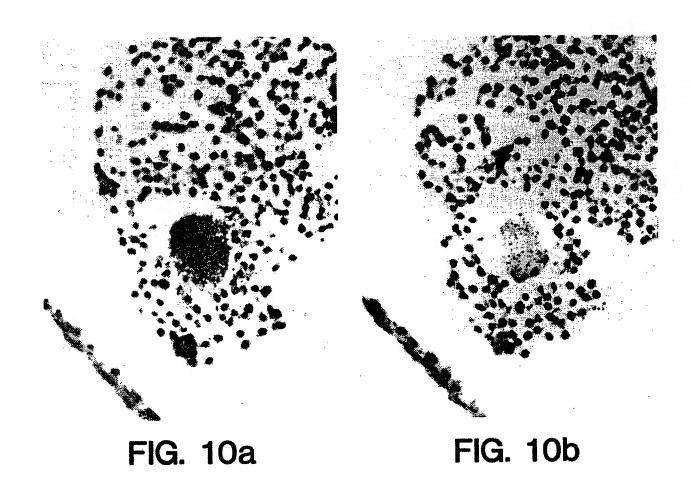


FIG. 8a

FIG. 8b



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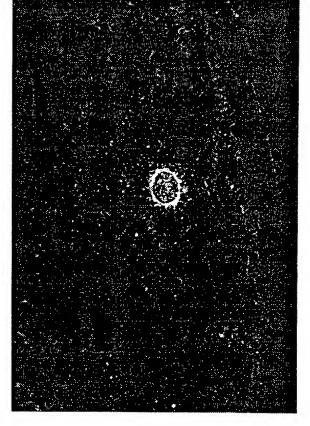


FIG. 11a

FIG. 11b

1						
US CL	US CL :Picase See Extra Sheet.					
According to International Patent Classification (IPC) or to both national classification and IPC  B. FIELDS SEARCHED						
	ocumentation searched (classification system followed	by classification symbols)				
	536/23.5, 23.4; 435/320.1, 69.1, 69.4, 91.1, 91.4, 2					
			in the fields seembed			
Documental	ion searched other than minimum documentation to the	exicni inai such documents are included	in the neigs searched			
1	ata base consulted during the international search (na		, search terms used)			
	Dialog, APS search terms: growth differentiation factor-9 Sequence search: GenBank, GeneSeq, PIR, SwissPro					
C. DOC	UMENTS CONSIDERED TO BE RELEVANT					
Category*	Citation of document, with indication, where ap	propriate, of the relevant passages	Relevant to claim No.			
Α	MOLECULAR ENDOCRINOLOGY,	· · · · · · · · · · · · · · · · · · ·	1-39			
	Lee, "Identification of a Novel Transforming Growth Factor-beta S					
	Transforming Growth Factor-beta Superfamily," pages 1034- 1039.					
	PROCEEDINGS OF THE NAT	IONAL ACADEMY OF	1-39			
Α	SCIENCES, USA, Volume 88,		1-33			
	"Expression of growth/differentiat	ion factor I in the nervous				
	system: Conservation of a bicis	. –	·			
	4250-4254, see entire document.					
X Furth	ner documents are listed in the continuation of Box C	. See patent family annex.	<u> </u>			
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	date and not in conflict with the application but cited to understand the					
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	cument published prior to the international filing date but later than priority date claimed	*&* document member of the same patent	t family			
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Box PCT	n, D.C. 20231	Shelly Guest Cermak	owar /			
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C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT					
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No			
X,P	THE JOURNAL OF BIOLOGICAL CHEMISTRY, Volume 268, Number 5, issued 15 February 1993, McPherron et al., "GDF-3 and GDF-9:Two New Members of the Transforming Growth Factor-beta Superfamily Containing a Novel Pattern of Cysteines", pages 3444-3449, see figure 2.	1-39			
	·				

A. CLASSIFICATION OF SUBJECT MATTER: US CL :				
536/23.5, 23.4; 435/320.1, 69.1, 69.4, 91.1, 91.4, 252.3, 252.33	; 530/350, 399, 388.23			
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